

Olive oil inhibits DMH-induced colon tumor growth by triggering mitochondrial apoptotic pathway: An *in vivo* study

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Abstract

Olive oil, a rich source of antioxidants, exhibits anti-inflammatory and anti-tumorigenic properties; however, the role of olive oil has not been systematically evaluated in the preclinical model of colon cancer. In this study, we investigated the chemopreventive role of olive oil on animal model of colon carcinogenesis by targeting its apoptotic machinery. In this study, DMH was used to induce colon cancer in rats; while olive oil was given to separate group of rats together with DMH treatment. Potent pro-apoptotic activity of olive oil was explored with apoptotic assays by FACS and confocal microscopy. It was observed that olive oil administration directly controls necrosis, and positively regulates apoptosis as assessed by Annexin V staining. Furthermore, ultrastructure examination by transmission electron microscopy (TEM) revealed that olive oil treatment ameliorates the mitochondrial damage in rat colon which was degenerated upon DMH treatment. Thus, these findings suggest that olive oil administration has a protective role in colon carcinogenesis and emphasizes new insights into chemoprevention by targeting mitochondrial/intrinsic cell death pathway.

Introduction

Colorectal cancer (CRC) is the third most common cancer and a major cause of morbidity and mortality. About 1.2 million new cases of colorectal cancer are diagnosed each year and more than 60 000 die due to this disease [1,2]. The continuing enormity of the colon cancer indicates that new approaches to the control of this malignant neoplastic disease are essentially required. Chemoprevention is a better approach to control incipient malignancy by regulating numerous intracellular-signaling cascades. Epidemiological studies links Mediterranean diet and cancer chemoprevention, revealed that the dietary factors significantly influence cancer risk in humans [3,4].

Olive oil is a key component of the Mediterranean diet and is believed to have numerous beneficial effects due to its antioxidant properties [4]. Olive oil is a chemopreventive phytochemical that can block the initiation process or reverse the promotion stage of multistep carcinogenesis. Several *in vitro* studies suggest that, in addition to antioxidant ability, olive oil phenols exert anti-inflammatory and anti-neoplastic activities against cancer cells [5]. In contrast, there are few reports in the literature that describe the effect of olive oil in cancer prevention and its underlying mechanisms, particularly in animal model of colon carcinogenesis. Therefore, additional investigations are required to evaluate the role of olive oil in colon cancer prevention, especially under *in vivo* conditions.

Thus, the present study was undertaken to investigate the potential chemopreventive efficacy of olive oil in rats subjected to DMH-induced colon cancer. It was observed that olive oil administration prevented tumor formation in the colon; the prevention is associated

with enhancement of apoptosis and reduction of necrosis. The study also explored the intrinsic/mitochondria-mediated apoptotic cell death pathway by examining the ultrastructure integrity of mitochondria to corroborate the potential chemopreventive effects of olive oil in colon carcinogenesis.

Materials and methods

Olive oil

Olive oil was purchased from company Figarro, Deoleo S.A., Spain. The nutrition facts of olive oil per 100g were: Energy 900Kcal, Protein 0g, Total carbohydrate 0g, Total fat 100g (Saturated fat 13g, Monosaturated fat 79g, Polyunsaturated fat 8g, Trans fat 0g), Cholesterol 0mg and Sodium 0mg.

Animals and experimental design

All animal studies were performed in accordance with protocols approved by the Institutional Animal Ethics Committee of PGIMER,

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Chandigarh. Healthy male Sprague Dawley rats weighing between 180 to 200g were divided into three groups having 24 rats in each group. Rats in Group I served as the normal control and received weekly subcutaneous (s.c.) injections of normal saline. Group II rats were injected subcutaneously with freshly prepared 1, 2 dimethylhydrazine (DMH) in normal saline, weekly at a dosage of 30mg/kg body weight, for two different time durations of 10 and 20 weeks. Rats belonging to Group III were treated with Olive oil at a dose of 1g/kg body weight thrice a week through oral gavage in addition to DMH treatment as was given to group II animals (Figure 1).

Transmission electron microscopy (TEM)

Tissues after removal from the rats were fixed immediately in 3% glutaraldehyde prepared in sorenson phosphate buffer for 24 hours, and subsequently were washed in Miellonings buffer and postfixed in 1% osmium tetroxide. The samples were then dehydrated at room temperature by keeping them in different grades of 100%, 90%, 70% and 50% ethanol for 3 minutes and three times, then infiltrated. Subsequently sections were embedded in 1:1 Propylene oxide/Epon resin and were then placed in an oven and allowed to polymerize at 60°C for at least 48 h. Semi-thin sections of thickness about 1µm were cut and stained with 0.5% toluidine blue prepared in 1% borax solution. Further, ultrathin sections of required area were cut by using an ultramicrotome and loaded on fine copper grids of 100-300 mesh size. The sections were double stained with uranyl acetate and lead citrate. These stained sections were finally viewed under Transmission Electron Microscope (Hitachi (H-7500) Chiyoda, Tokyo, Japan), operating at 40-120 kV [6].

Apoptosis assay

Confocal microscopy: The colonic tissues were cut into 10µm sections by using cryostat. Thereafter, the sections were washed with PBS buffer, incubated for 15 min with AnxV-FITC, and washed with Annexin binding buffer, and then incubated with Propidium iodide by using Annexin V-FITC Apoptosis Detection kit (Calbiochem). The

sections were mounted with anti-fading agent and examined under a confocal microscope (Olympus 1x81) using standard settings for FITC fluorescence-detection [7].

Flow cytometry: For flow cytometry, colonic tissues were dissected and minced to small clumps, followed by enzymatic dissociation with 0.5mg collagenase IV (Sigma) in PBS for 25 min at 37°C on a shaker and centrifuged at 500xg for 10 min. The supernatant containing debris was discarded; the pelleted cells were washed twice with PBS and filtered through a 100µm cell strainer to obtain single-cell suspension. Dissociated colonic cells were finally resuspended in PBS. Furthermore, cells were used for apoptotic assays using Annexin V-FITC Apoptosis Detection kit (Calbiochem) according to manufacturer's instructions. Cell sorting was performed on a BD FACS Aria II instrument in the Flow Cytometry [7].

Results

Olive oil treatment stimulates apoptosis and prevents necrosis

To examine, if olive oil, modulates the apoptotic pathway, we examined the apoptotic rate of colonic cells by using annexin V staining. The results were demonstrated by confocal microscopy and further validated by FACS. Interestingly, the results revealed more necrosis and less apoptosis with DMH treatment as compared to normal animals. However, olive oil treatment caused an appreciable decrease in necrosis and an increase in early and late apoptosis in DMH-treated rats.

Confocal microscopy

Based on the principle of this technique, the normal cells were not stained by the two dyes (Annexin V FITC-/PI-); the early apoptotic cells could only be stained by Annexin V-FITC (Annexin V-FITC+/PI-); the necrotic cells were only stained with PI (Annexin V-FITC-/PI+); the late apoptotic cells were positive in both Annexin V-FITC and PI staining (Annexin V-FITC+/PI+). AnxV-FITC (green) and PI (red) stains were applied to cryostat sections of colonic tissues of all the treated and normal control rats. As shown in (Figure 2). control cells were more stained with AnxV-FITC, indicating the higher apoptotic rate in the normal control group. In the colonic sections treated with DMH alone for 10 or 20 wks, we observed less stain by Annexin

V-FITC (green), indicating that apoptotic rate was much less in colonic tissues of DMH alone treated rats. Also, we observed more PI stain (red) which confirmed that more cells were in the necrotic stage, after DMH alone treatment. However, colonic sections of DMH+olive oil treated rats at 10 or 20 weeks indicated that most cells were stained by Annexin V-FITC (green), but less by PI, indicating that most of the olive oil -treated cells were in the early apoptotic stage. Also, some other cells were stained by both the dyes, demonstrating that these cells were in the late apoptotic stage in these tissues (Figure 2).

FACS analysis

Similarly, these results also revealed that control cells were more stained with Annexin-V-positive (7.4%), Annexin-V/PI-positive (18.1%) and less stained with PI-positive (14.4%), indicating the higher apoptotic rate in the normal control group. However, there were very few Annexin-V-positive (3.1%), AnnexinV/PI-positive (7.45%) and more PI-positive cells (22.8%) in the DMH-treated group at 10 weeks. However, less than 20% PI positive cells were observed in olive oil treated group. While greater than 25% positive cells were noted for both Annexin V-FITC/PI staining at 10 weeks with olive oil treatment.

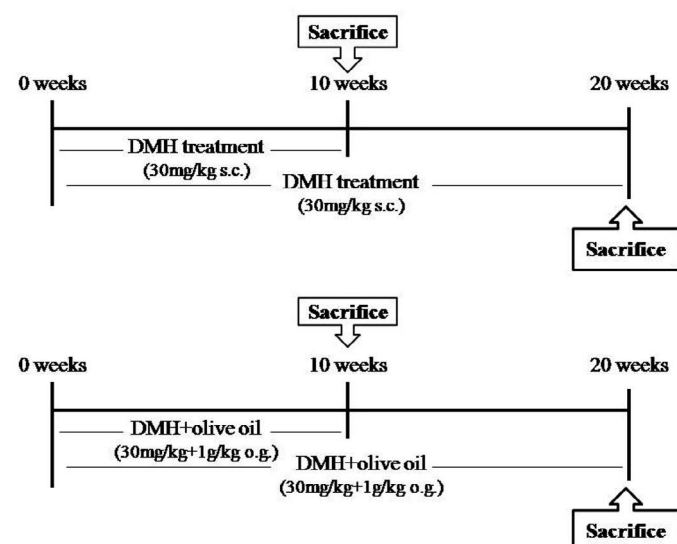


Figure 1. Olive oil treatment induces apoptosis and decreases necrosis. Apoptosis/Necrosis detection in control and treated rats by Confocal microscopy. Annexin V-FITC/PI staining was used to observe the induction of apoptosis. Annexin V stained the membrane of apoptotic cells (green fluorescence), Propidium iodide (PI) stained the nucleoli of necrotic or dead cells (red fluorescence) and merged of Annexin V-FITC/PI stained the late apoptotic cells (dual channel fluorescence). Magnification ×200

Annexin V-FITC/PI staining was also completed for the 20 weeks DMH-alone and DMH +olive-treated cells. 0.4% and 21.55% annexin V positive cells; 48.4% and 9.9% PI positive cells observed for DMH-alone and DMH+olive oil-treated cells at 20 weeks, respectively. While greater than 40% positive cells were noted for both Annexin V-FITC/PI staining at 20 weeks with olive oil treatment as compared to DMH-alone treatment (Figure 3).

Olive oil restores the mitochondrial integrity in colon cancer

Mitochondria is a central player controlling cell death, triggers the activation of caspase cascade, leading to intrinsic/mitochondrial-mediated apoptotic pathway. Thus, in the present study, we also examined the viability and morphological changes in the mitochondria from colonic tissues of treated and normal control rats under transmission electron microscope. Ultrastructure examination of colonic sections of normal controls, revealed good preservation of mitochondrial morphology and integrity, while rats treated with DMH-alone at 10 or 20 weeks exhibited mitochondrial damage with alteration in size, disappeared matrix, disorganized cristae and in some mitochondria only the outer membrane remained. However, at 10 or 20 weeks, olive oil administration protected the mitochondria by reducing the mitochondrial injury and restoring its integrity (Figure 4).

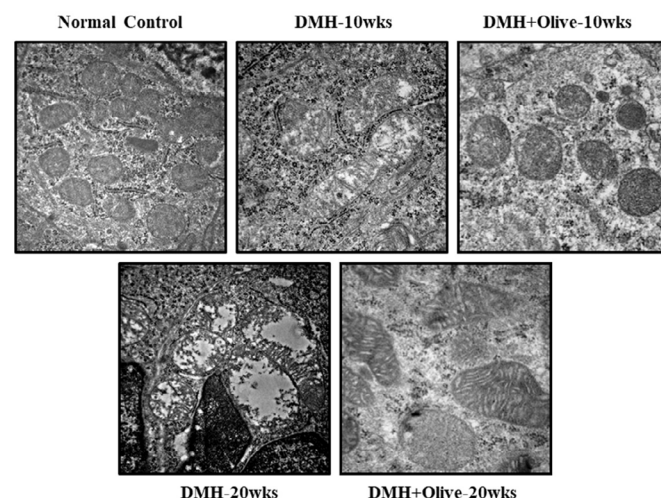


Figure 4. Olive oil administration protected the mitochondria by reducing the mitochondrial injury and restoring its integrity

Discussion

Despite persistent focus on early detection and treatment of cancer, many develop disease due to less prominence on prevention. Thus, in the present study, with more emphasis on prevention, role of olive oil and its potential has been successfully elucidated in the CRC pathogenesis with a special focus on the molecular basis of its heterogeneity. It is now very well evident that dietary agents inhibit tumor growth by targeting different mechanisms involved in the process of carcinogenesis that further leads to the induction of apoptosis [8]. Similarly, we also confirmed the direct effect of olive oil on the rate of apoptosis/necrosis that showed a decrease in necrosis and increased early/late apoptosis in comparison to DMH treatment as shown by annexin V staining. It is believed that necrosis might be considered as a prognostic factor of colorectal cancer, but the underlying mechanism remains unclear [9-11]. Since TNF (tumor necrosis factor) is secreted by inflammatory cells, may be involved in inducing necrosis and stimulates tumor promotion, thus, the plausible reason for this may be TNF- α -induced NF- κ B activation resulted in an inflammation mediated carcinogenesis [12-14]. However, in cancer, TNF is a double-edged sword which could be either pro- or anti-tumorigenic. On one hand, TNF could be an endogenous tumor promoter, because TNF stimulates the tumor promotion, angiogenesis, invasion and metastasis of cancer cells via its molecular targets NF- κ B, VEGF and MMPs [15-18]. On the other hand, TNF could be a cancer killer by JNK activation, an important contributor to death response via inhibition of NF- κ B activation which further increases susceptibility to TNF-induced death [19]. JNK activation is required for cytochrome c release and further activation of caspase cascade to induce apoptosis [20]. Pham et al., (2004) demonstrated that activation of NF- κ B inhibits TNF- α -induced apoptosis by controlling the activation of JNK cascade. Thus, the present results clearly demonstrated that olive oil may regulates apoptosis/necrosis by activation of caspase cascade and inhibition of TNF- α -induced NF- κ B pathway. Our findings gain strength from the study of [21], who elucidated two mechanisms of apoptosis either by p53 mediated mitochondrial pathway that leads to the activation of caspase-9 which in turn activates caspase-3 [22] or by NF- κ B-mediated pathway. This supports the present findings involving the chemopreventive role of olive oil in this process.

After observing appreciable effect of olive oil on rate of apoptosis and necrosis, we assumed that olive oil induced apoptosis is supposed to

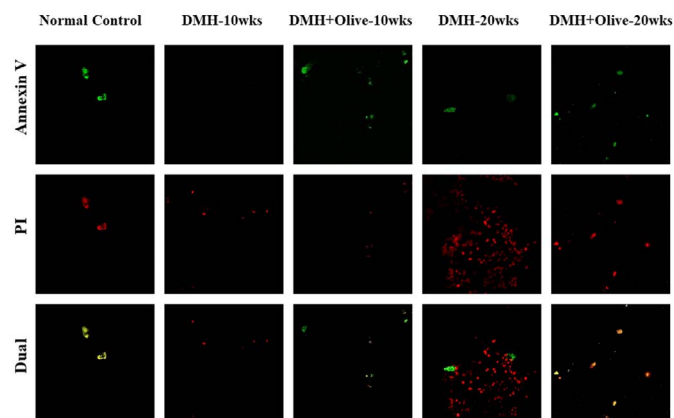


Figure 2. Apoptosis/Necrosis detection by FACS analysis. The induction of apoptosis was determined by flow cytometric analysis of Annexin V-FITC and PI-staining. Cells in the lower right quadrant (Q4) indicate Annexin V-positive (early apoptotic), upper right quadrant (Q2) indicate Annexin V-positive/PI-positive (late apoptotic), upper left quadrant (Q1) indicate PI-positive (necrotic/dead), and lower left quadrant (Q3) indicate live cells

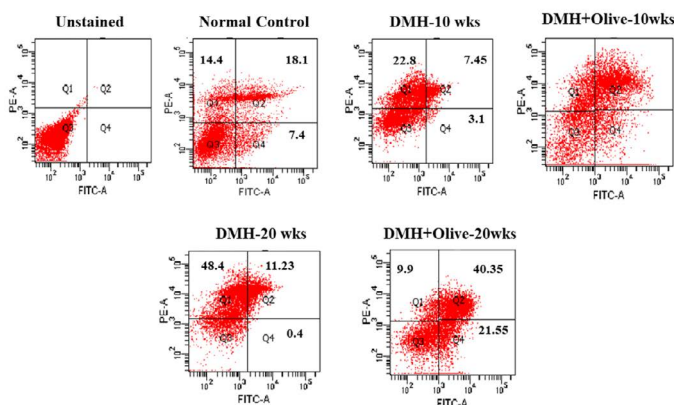


Figure 3. Effect of olive oil on mitochondria at ultrastructural level using transmission electron microscopy. Electron micrograph of normal control and treated segments of colon, Scale bar 500nm

be due to the activation of mitochondrial/intrinsic apoptotic pathway. Thus, we desired to observe the effect of olive oil on mitochondrial morphology to substantiate the above presumption. In the present study, the viability and morphological changes in the mitochondria from colonic tissues were determined by transmission electron microscopy.

Mitochondrial dynamics have roles beyond maintenance of morphology, had impact on cell death, and its alterations are important for inflammatory bowel disease, colitis and colon cancer patients [23-25] observed the mitochondria hyperplasia and absence of goblet cells in neoplastic glands of colonic tissue in DMH-induced rat model. Other studies have shown that mitochondrial numbers are varied in DMH-treated animals as well as human biopsies of colon cancer as compared to control counterparts [26,27]. Likewise, we also observed the mitochondrial alterations in DMH-treated rat colon, while it has been demonstrated that olive oil treatment ameliorates the mitochondrial damage in rat colon that was degenerated upon DMH treatment [28-30]. As of now, no information is available with regard to ultrastructure examination of colonic tissues inflicted with DMH injury and co-treatment with olive oil. Since, [24] showed that mitochondria are dynamic structures that undergo fusion and fission process continuously throughout the life of a cell, possibly in present study, olive oil keep the balance between fission-fusion process of mitochondria to maintain its functional level. However, further studies may be needed to explore the dynamic structure of mitochondria after olive oil treatment, that may further explain the chemopreventive role of olive oil in colon carcinogenesis.

Conclusion

The present study revealed the potential role of olive oil in preventing the initiation and development of colon carcinogenesis in experimental animals. The mode of action of olive oil may involve activation of mitochondrial/intrinsic apoptotic pathway characterized by alteration of mitochondrial structure and rate of apoptosis, which may have associated with the suppression of inflammatory pathway. Thus, the current findings might build a persuasive case for chemopreventive role of olive oil in targeting the different stages of colon carcinogenesis and open up a new vista for future research to be translated to human subjects to control incipient malignancy.

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Declarations of interest

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