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Standardizing transdifferentiation of rabbit bone marrow mesenchymal stem cells to corneal lineage by simulating corneo-limbal cues

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Corneal pathologies due to limbal stem cell deficiency or trauma is a common occurrence with high incidence in the modern era. Absence of native limbal stem cells in these cases can even result in blindness unless treated. Identifying alternate autologous stem cells with potential to replace limbal stem cells, gain its significance especially in cases of bilateral limbal damage. Here we look at the suitability of proposing bone marrow derived mesenchymal stem cells as an alternate to limbal stem cells. This work had made use of the corneo-limbal cues for trans differentiating these mesodermal cells to corneal epithelial lineage. Mesenchymal stem cells were isolated from rabbit bone marrow and the culture conditions were optimized. Characterized mesenchymal stem cells were transdifferentiated by simulating limbal niche using extracellular matrix and limbal fibroblast conditioned medium. The differentiation to corneal lineage was evaluated by analyzing the expression of mesenchymal (ABCG2, CD29) and corneal epithelial (CK3/12) markers. Cells cultured in the presence of conditioned medium alone and with conditioned medium/extracellular matrix showed the expression of CK3/12. CK3/12 expressing cells in these cultures were increasing, with a concomitant decrease in stem cell marker expression. Trans differentiating mesenchymal stem cells to corneal epithelial like cells will provide a useful, autologous source of adult stem cells in therapeutic approaches for patients suffering from bilateral limbal stem cell deficiency.

Introduction

Visual loss is increasing in an alarmingly high rate across the globe with profound psychological, economic, social and financial consequences for the patient and the society. Cornea, the outer most layer of the eye prevents foreign body entry, protects the eye and supports vision. Cornea is made of five different layers with corneal epithelium replenished regularly by fresh epithelial cells derived from limbal stem cells (LSCs) [1]. Limbal stem cells are located within the basal layer of corneo-scleral junction called limbus [1,2]. Deficiency of LSCs due to inherited and acquired conditions prevents the re-epithelialization of cornea by limbal epithelial cells resulting in conjunctivalization and vascularization that leads to visual impairment or blindness. Many efforts are made to improve the clinical prognosis and treatment for limbal stem cell deficiency (LSCD) like autologous cultured limbal epithelial transplantation (CLET), which requires limbal biopsy from contra lateral eye, which can also lead to LSCD in the normal eye. However such treatment modalities are not possible in patients with bilateral total limbal stem cell deficiency conditions. In bilateral cases, allogeneic limbal epithelium harvested either from living related donors or cadaveric donor eyes may be used for transplantation in combination with long term immunosuppressive therapy. This approach has a very low success rate as compared with autologous cells mostly followed by implant rejection. Therefore, alternative stem cell-based therapeutic strategies for ocular surface reconstruction are required and is in focus. *Ex vivo* expansion and delivery of alternate autologous adult stem cells like oral mucosal epithelium [3-5], conjunctival epithelium [6,7] and hair follicle derived stem cells [8] are now being looked into as promising approach for

LSCD treatment. There are scanty reports on the use of such alternate cell sources and the clinical outcomes are often less satisfactory [9,10]. The success of treatment methodology involving *ex vivo* expansion and transplantation largely depends upon the stabilization of physiologic conditions of the eye and in providing a niche for the restoration of stem cell milieu and graft survival.

It is well established that epithelial-mesenchymal interactions of eye plays a major role in wound healing, development and maintenance of LSCs. Mesenchymal stem cells (MSC) due to its multifunctional properties like immunomodulatory and anti-inflammatory functions [11], multipotency [12,13] and relative ease of expansion in culture represents a readily accessible source of adult cells for tissue repair and reconstruction. Moreover, promising results has been obtained in recent studies with MSC for treating corneal chemical burns [14-17]. In this context, it would be very much appreciable to transdifferentiate MSC to corneal epithelial like cells prior to transplantation, which could be used as an alternate autologous cell source in treatments involving bilateral limbal deficiency. This study was aimed in transdifferentiating MSC to corneal epithelial like cells *in vitro* by simulating corneo-limbal niche using extracellular matrix (ECM) and conditioned media.

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Materials and methods

Isolation and maintenance of rabbit bone marrow mesenchymal stem cells

MSC were isolated from the femurs of two-year-old New Zealand White rabbits by direct plating method as previously described [18]. The study was approved by the Institute Animal Ethics Committee (IAEC) of Sree Chitra Thirunal Institute of Medical Sciences and Technology (SCTIMST). Briefly, the femur was collected in phosphate buffered saline (PBS) containing 1X antibiotic (penicillin /streptomycin, PAA laboratories, Germany). The bone was cleaned and bone marrow was flushed out. The cell suspension was mixed, centrifuged at 600 g for 10 min (Eppendorf, Germany) and was resuspended in Dulbecco's Modified Eagle Medium (DMEM) (Himedia, India) containing 10% fetal bovine serum (FBS) (Lonza, USA). Isolated cells were cultured on 25 cm² culture flask (Cell Star, Germany) and maintained at 37°C, 5% CO₂, 95% humidity in a CO₂ incubator (Sanyo, Japan). Medium change was given thrice a week.

Proliferation of rabbit mesenchymal stem cells

Proliferation of MSC in three different commercially available media: DMEM, DMEM low glucose (DMEM LG) and α-minimum essential medium (α MEM) (Invitrogen, USA) were analyzed. After attaining confluence, MSC were seeded at a density of 2 × 10³ cells/ cm² on 12 well plate (Cell Star, Germany) and were cultured in the respective medium for 24 h and 48 h. At the end of culture period, proliferation of MSC was determined by MTT [(3-(4,5- Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] assay. Cells were treated with MTT (Sigma Aldrich, Germany) (0.5 mg/ml) for 4 h to allow the formation of formazan crystals, followed by their solubilization with isopropanol (MERCK, USA). Supernatant was transferred to a 96 well plate and the absorbance was read using a spectrophotometer (Bioteck Powerwave XS, USA) at 570 nm. Optimum medium was selected for further experiments and MSC cultured under this condition was analyzed by immunocytochemistry and flow cytometry for the expression of mesenchymal lineage specific markers (CD105, vimentin, ABCG2, CD90 and CD29).

Isolation and culture of rabbit limbal fibroblasts

From the harvested eyes, the limbus was excised and washed twice in PBS containing 1X antibiotic and de-epithelialized by treating with 2 mg/ml dispase II (Roche, USA) for 20 min followed by mechanical debridement. Stromal layer was excised off from de-epithelialized limbal tissue and was incubated for 2 h in collagenase - type I (1 mg/ml) (Sigma Aldrich, Germany), at 37°C in a CO₂ incubator. The resultant digest was centrifuged and the pellet was resuspended in DMEM (Invitrogen, USA) containing 10% FBS. The cells were cultured in 60 mm dishes (Cell Star, Germany) for 21 days and the cell growth were monitored using phase contrast microscope (Leica, Germany). Cultured limbal fibroblasts (LF) were characterized by immunocytochemistry and flow cytometry for the expression of fibroblast lineage specific markers Secreted Protein Acidic and Rich in Cysteine (SPARC), vimentin and CD90.

Transdifferentiation of MSC to corneal lineage

MSCs were differentiated to corneal lineage by simulating the corneo-limbal niche factors using extracellular matrix (ECM) coated plates/dishes/coverslips (CVs) and limbal fibroblast conditioned medium (CM).

Coating of extracellular matrix components

Type I collagen (Nutacon, Netherlands) was used as the ECM component for transdifferentiation studies. Collagen was coated by adding collagen solution (1.5 mg/ml) to dish and spreading uniformly. Plates were then incubated for 1 h at 37°C and were gently washed with PBS before use.

Preparation of conditioned medium

Conditioned medium was prepared using the spent medium of the LF primary cultures. The spent medium was collected on every alternate day from LF cultures, centrifuged at 3000 rpm for 15 min to remove cell debris, filtered using 0.22 micron syringe filter, aliquoted and stored at -80°C until use.

Simulation of niche factors for inducing transdifferentiation of MSC

To study the role of niche factors in the differentiation of MSC, three different culture systems were selected (CS_{ecm}, CS_{ecm+cm} and CS_{cm}). A combination of ECM and CM was used to formulate this culture system [(CS_{ecm}: ECM coating alone), (CS_{ecm+cm}: ECM coating + CM), (CS_{cm}: CM alone)]. MSC were cultured in this culture system for 7 days and the transdifferentiation to corneal epithelial lineage was analyzed by immunocytochemistry and flow cytometry for the expression of ABCG2, CD29 and cytokeratins 3/12. RT-PCR was also done for evaluating CK12 expression.

Immunocytochemistry

MSC, LF and transdifferentiated MSC were characterized by immunocytochemistry. Cells cultured on CVs were fixed using 4% buffered paraformaldehyde (30 min) and permeabilized with 0.1% Triton X solution (Sigma Aldrich, Germany). Nonspecific binding of antibodies was blocked by incubating cells in 1% bovine serum albumin (BSA) for 10 min. Cells were then rinsed and incubated with primary antibody (1:100) (table-1) for 1 h and was followed by incubation with goat anti mouse IgG-FITC (Sigma Aldrich, Germany) (1:100) for 1 h in dark. Cells were rinsed and counter stained with propidium Iodide (PI) (Sigma Aldrich, Germany) (0.05 µg/ml) for 2 min. CVs were then mounted with fluorescent mounting medium (Dako Cytomation, USA) and was analyzed by Laser Scanning Confocal Microscope (LSM510 META, Carl Zeiss, Germany) using Argon and HeNe lasers. Fluorescence emission was detected using filter of FITC and PI.

Flow cytometry analysis

MSC, LF and trans differentiated MSC were further characterized by flow cytometry analysis. Cells were harvested by trypsinization and made into single cell suspension by passing through a cell strainer of pore size 70 µm (BD falcon, USA). The cell suspension was pelleted by centrifuging at 600 g for 30 min, fixed in 4% buffered paraformaldehyde, repelled and permeabilized using 0.1% Triton X. The nonspecific binding sites were blocked by incubating with 1% BSA and cells were incubated with primary antibody (1:100) (Table 1) for 45 min at room temperature. Primary antibody was replaced with appropriate secondary antibody (1:100) and incubated further for 45 min. The cells were then pelleted and resuspended in PBS for analysis using a flow cytometer (FACS Aria, BD USA). The results were analyzed by BD FACS Diva software.

RT-PCR analysis

Transdifferentiated MSCs were characterized by RT-PCR as described previously [19]. RNA was collected from cells on 3rd and 7th

Table 1. Antibodies used for immunostaining / flow cytometry.

Primary antibody	Characteristic protein	Company
Monoclonal mouse anti cow vimentin clone (Vim 3B4)	Intermediate filaments, vimentin	Dako Cytomation, USA
Mouse anti human integrin $\beta 1$ four (CD29)	Integrin	Millipore, USA
FITC mouse anti human CD90	Thymine 1	BD Pharmingen, USA
ABCG2 (BRCP1) purified mouse anti human monoclonal antibody	Transmembrane protein	BD Pharmingen, USA
Endoglin rat monoclonal IgG _{2a} CD105	TGF β receptor	Santacruz Biotechnology, USA
Osteonectin /SPARC	Bovine bone osteonectin	QED bioscience, USA
Monoclonal mouse anti human cytokeratin clone AE1/AE3,	cytoskeleton	Dako cytation, USA

All antibodies were used in 1:100 dilutions.

day by treating with TRIzol reagent (Bangalore Genei, India). RNA was isolated, precipitated and resuspended in nuclease free water. Complementary DNA (cDNA) was synthesized from 1.2 μ g RNA using M-MuLV RT-PCR kit (Bangalore Genei, India) in a thermocycler (Eppendorf AG, Germany). PCR was then performed using Genei Red Dye PCR Master Mix (Bangalore Genei, India) according to manufacturer's protocol. PCR amplification was performed with specific primer pairs for different markers [20], as given in table 2. The cyclic conditions were set for 35 cycles, each with denaturation at 94°C for 30s, specific annealing temperature (Table 2) for 30s and extension at 72°C for 1 min. The PCR products were subjected to electrophoresis in 2% agarose gel containing 0.4 μ g ethidium bromide (Bangalore Genei, India) and was imaged using a Phosphor Imager (Fujifilm FLA 5100, Japan).

Statistical analysis

Data obtained from three independent experiments were expressed as mean \pm standard deviation and was used for the analysis. Statistical significance between groups was analyzed using student's t test. For all analysis, differences were considered significant when $p < 0.05$.

Results

Culture and characterization of MSC

MSC were isolated and cultured based on their ability to adhere to plastic cell culture plates. These adherent cells were spindle-shaped in morphology and proliferated as colonies. Optimization of culture conditions for *in vitro* expansion and maintenance of MSC was done by assessing their proliferation by MTT assay. Among the three media screened, α MEM was found to be supporting more proliferation compared to DMEM-LG and DMEM ($p < 0.001$) (Figure 1). Hence α MEM was selected for culturing MSC for further experiments.

The immunocytochemistry of MSC cultured in α MEM showed expression of CD105, ABCG2 and vimentin, but was negative for CD90 (Figure 2a-d). These findings were further confirmed by flow cytometry. Flow cytometry analysis showed cultured MSC were positive for the expression of vimentin ($99\% \pm 0.44\%$), CD105 ($99\% \pm 0.54\%$) and ABCG2 ($99\% \pm 0.63\%$). The cultured MSC showed reduced expression of CD29 ($75\% \pm 5.63\%$) and negative expression of CD90 ($1\% \pm 0.11\%$) (Figure 2e-j).

Culture and characterization of limbal fibroblast

LF isolated from de-epithelialized limbal tissue cultured in DMEM were characterized for limbal fibroblasts specific markers (SPARC, vimentin and CD90) by immunocytochemistry and flow cytometry.

The cultured cells showed typical spindle morphology of fibroblasts with extensive proliferation (Figure 3a-c). Immunocytochemistry showed their cytoplasmic expression of SPARC and vimentin, but were negative for CD90 expression (Figure 3d-f). Further quantitative analysis by flow cytometry reconfirmed the immunocytochemistry data (vimentin = 98%, SPARC = 99% and CD90 = 0.3%) (Figure 3g-j). These results confirmed that the cultured cells from de-epithelialized limbal tissue were LF.

Transdifferentiation of MSC to corneal lineage

The role of limbal niche factors [ECM and LF released factors (CM)] in inducing transdifferentiation of MSC to corneal lineage was evaluated using three culture systems (CS_{ecm} , CS_{ecm+cm} and CS_{cm}) by monitoring expression of corneal epithelial markers (CK3/12) and stem cells markers (ABCG2 and CD29) on 3rd and 7th day of culture. Immunocytochemistry showed the expression of ABCG2 by cells cultured in CS_{ecm} , but were negative for CK3/12 on both 3rd and 7th day (figure 4a-d). In CS_{ecm+cm} and CS_{cm} , cells expressed both ABCG2 and CK3/12 (figure 4e-l). Further analysis of cellular gene expression profile for CK12 showed their negative expression in CS_{ecm} , but was positive in CS_{ecm+cm} and CS_{cm} , on both 3rd and 7th day (Figure 4m). This result was further evaluated and quantified by flow cytometry for CS_{ecm} , (Figure 5), CS_{ecm+cm} (Figure 6), and CS_{cm} (Figure 7). The quantitative values obtained for the expression of these markers in three culture systems is summarized in table 3. Flow cytometry data reconfirmed the findings obtained by immunocytochemistry and RT-PCR.

Discussion

Corneal disorders are the fourth leading cause of blindness and a major problem in ophthalmology [21]. This emphasizes the need for continual search for improved treatment regimes. Corneal integrity and function is dependent on the homeostasis of the outermost corneal layer, the corneal epithelium. Corneal epithelium is replenished at definite intervals by LSCs located in the basal layer of the limbus in

Table 2. Primer sequence, annealing temperature and product length.

Gene	Primer sequence	Annealing Temperature (°C)	Product length (bp)
B Actin	FP- ATCGTGTGGACTCCGGCGA RP- AGGAAGGAGGGCTGAAACAG	52	350
CK 3	FP- GGCAGAGATCGAGGGTCTC RP- GTCATCCTCGCTGCTGTAG	64	145
CK 12	FP- CATGAAGAAGAACACGAGGATG RP- TCTGCTCAGCGATGGTTCA	63	150

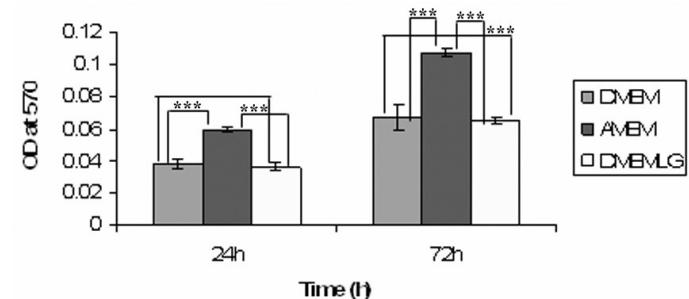


Figure 1. Proliferation of mesenchymal stem cells. MTT assay profile of mesenchymal stem cells cultured in different media after 24h and 72h. Data expressed as mean \pm SD of three different experiments. Cellular activity of MSC cultured in α -minimum essential medium (α MEM) was compared with that of Dulbecco's Modified Eagle Medium (DMEM) and Dulbecco's Modified Eagle Medium-low glucose (DMEM LG) and significant differences ($p < 0.05$) are denoted by **.

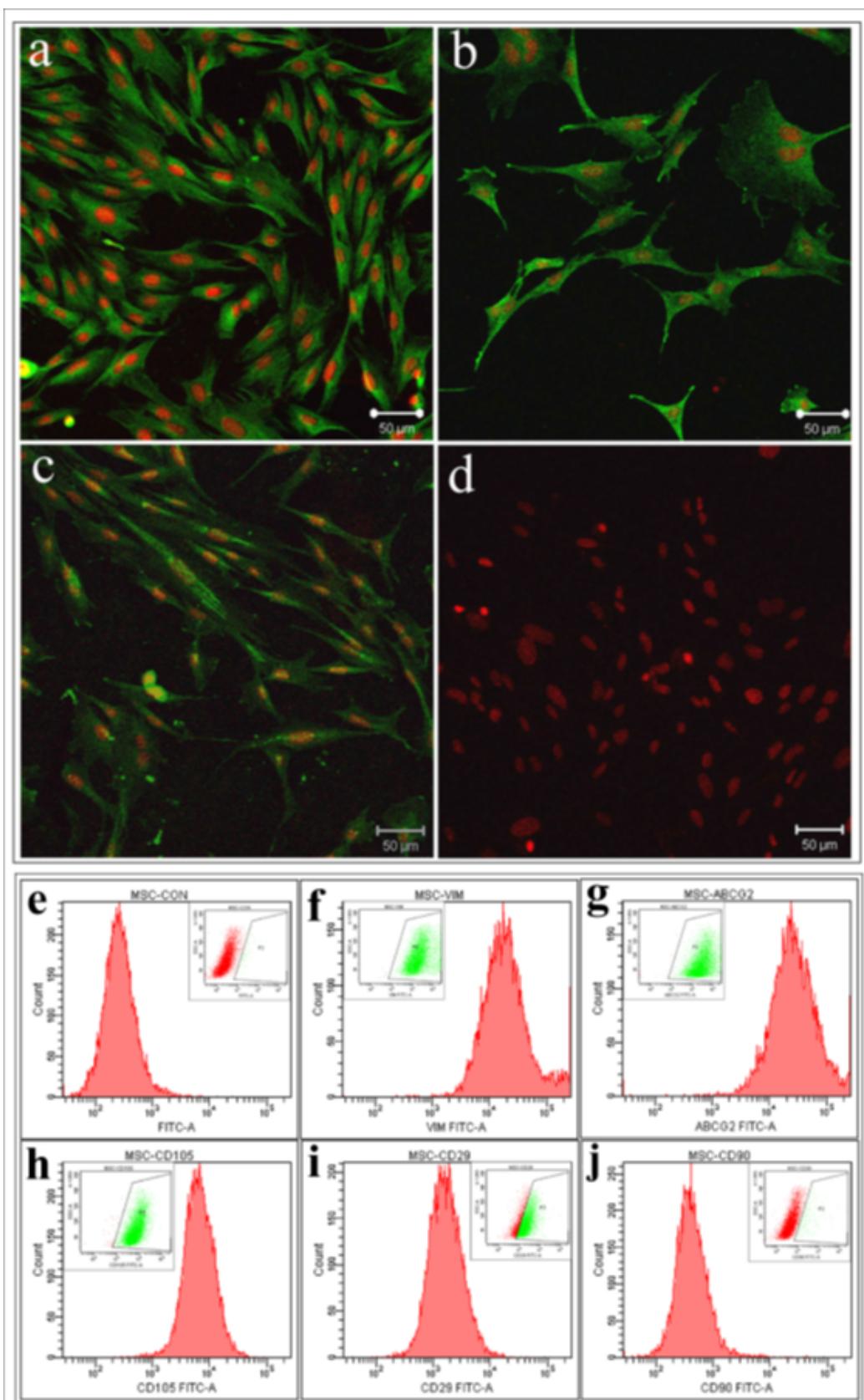


Figure 2. Characterization of mesenchymal stem cells: Representative confocal image showing the expression of (a) Vimentin, (b) ABCG2, (c) CD105 and (d) negative expression of CD90 in rabbit bone marrow mesenchymal stem cells. Scale bar denotes 50 μm. Flow cytometry analysis of rabbit bone marrow mesenchymal stem cells showing the expression of (f) vimentin (g) ABCG2 (h) CD105 (i) CD29 and (j) negative expression of CD90. The inset image denotes the gated population and (e) represents the isotype control.

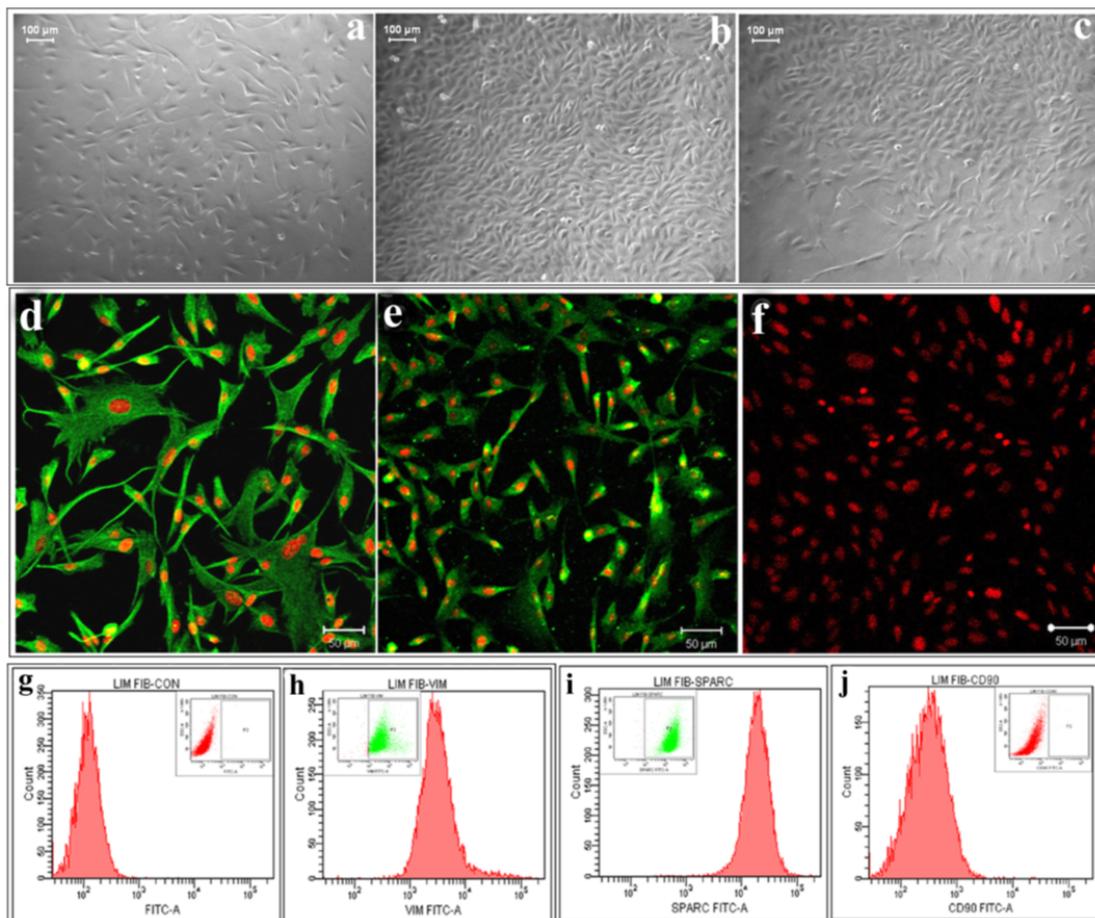


Figure 3. Characterization of LF: phase contrast image of LF in primary culture showing elongated fibroblast morphology on (a) 4th day, (b) 10th day and (c) after passaging. Immunocytochemistry showed the expression of (d) vimentin, (e) SPARC and (f) negative expression of CD90 in LF cells. Scale bar denotes 50μm. Flow cytometry analysis of LF cells showing the expression of (h) vimentin (i) SPARC and (j) negative expression of CD90. The inset image denotes the gated population and (g) represents the isotype control.

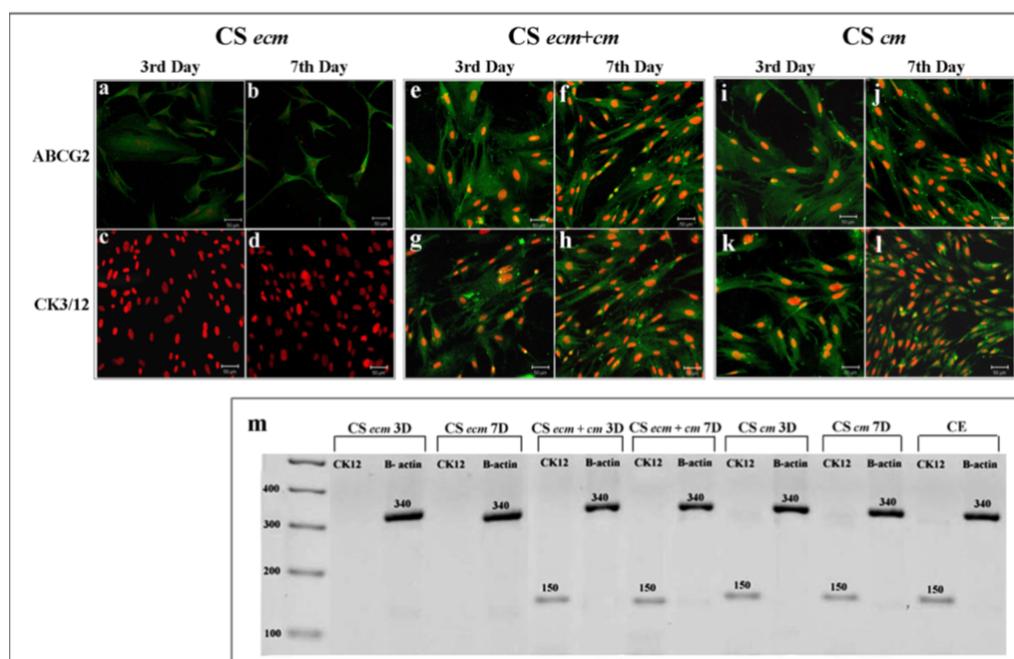


Figure 4. Transdifferentiation of MSC in culture systems. Immunocytochemistry of cells in CS_{ecm} (a-d), CS_{ecm+cm} (e-h) and CS_{cm} (i-l) on 3rd and 7th day showing the expression of ABCG2 and CK3/12. Scale bar denotes 50μm. (m) RT PCR analysis showing the gene expression of CK12 in culture systems on 3rd and 7th day. The value above the band denotes the product length. Abbreviations: CS_{ecm} 3D: CS_{ecm} 3rd day; CS_{ecm} 7D: CS_{ecm} 7th day; CS_{ecm} 3D: CS_{ecm} 3rd day; CS_{ecm+cm} 3D: CS_{ecm+cm} 3rd day; CS_{ecm+cm} 7D: CS_{ecm+cm} 7th day; CS_{cm} 3D: CS_{cm} 3rd day; CS_{cm} 7D: CS_{cm} 7th day; CE: corneal epithelium; CK12: cytokeratin 12; B actin: β actin.

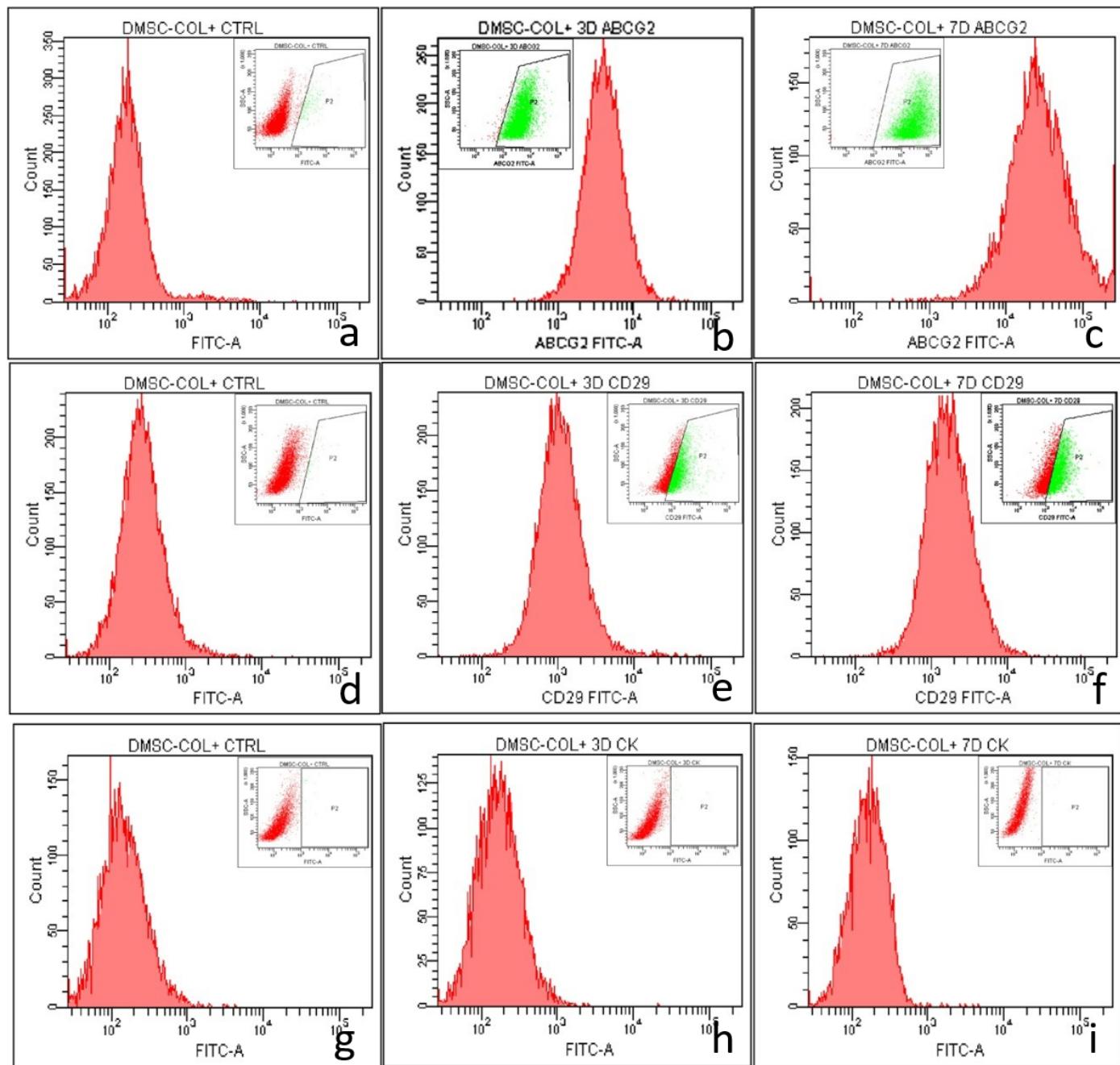


Figure 5. Flow cytometry analysis of MSC in CS_{cem} showing the expression of (a, b, c) ABCG2 (a) control (b) 3 day (c) 7 day, (d, e, f) CD29 (d) control (e) 3 day (f) 7 day and (g, h, i) CK3/12 (g) control (h) 3 day (i) 7 day. The inset image denotes the gated population.

close proximity to limbal niche [1,2], which plays an important role in the maintenance of stem cells [22]. Any damage or dysfunction of this limbal niche can affect the LSC populations and can severely compromise ocular surface integrity and vision [23].

Selection of treatment regime depends on the extent of damage to the limbal niche and the LSCs. In severe cases a cell transplantation therapy is necessary where the host stem cells are inefficient to revive the corneal layer. In order to replenish the stem cell population in eye, it is always desirable to use autologous cells for transplantation since it avoids the risk of immune rejection and the need for immunosuppression. However in bilateral LSCD condition, transplantation of allogenic stem

cells or *ex vivo* expanded cells are employed due to lack of autologous LSCs cell sources. Allograft transplantation requires long-term immunosuppression that involves high risks of systemic complications. A major strategy for bilateral LSCD treatment involves the *ex vivo* expansion, differentiation and delivery of alternate autologous stem cells *viz* delivery systems like amniotic membranes and fibrin glue [24]. Identifying the best non-ocular cell source is now well attempted in the field of ophthalmology and bone marrow stem cell is one prominent group in that research stream. Bone marrow contains hematopoietic and mesodermal precursors. MSC are of intense therapeutic interest because of its multipotency, ready availability, relative ease of expansion in culture and low immunogenicity [25-27]. Recently, there have

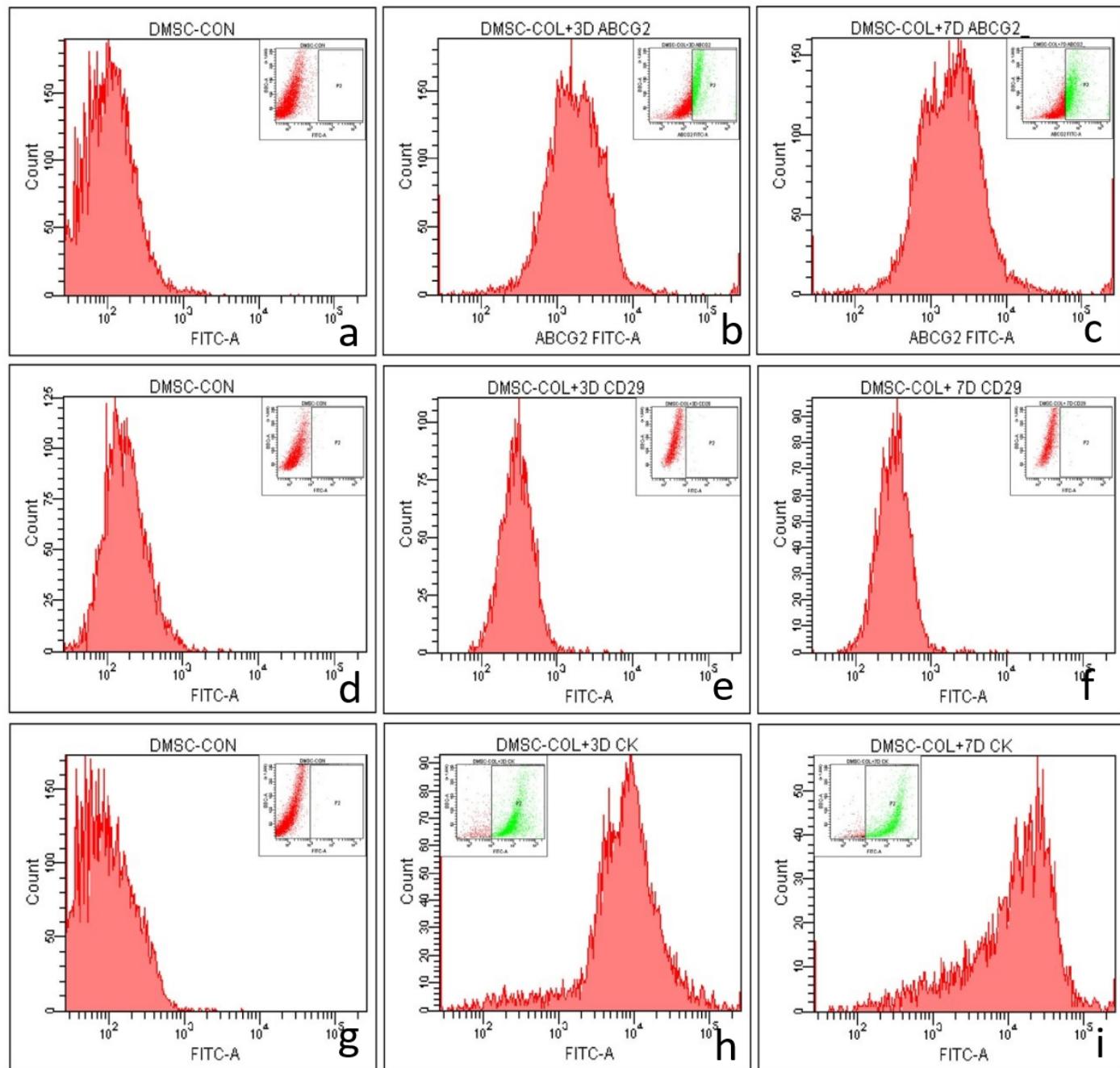


Figure 6. Flow cytometry analysis of MSC in CS_{ecm+cm} showing the expression of (a, b, c) ABCG2 (a) control (b) 3 day (c) 7 day, (d, e, f) CD29 (d) control (e) 3 day (f) 7 day and (g, h, i) CK3/12 (g) control (h) 3 day (i) 7 day. The inset image denotes the gated population.

also been significant advances in the use of autologous mesenchymal stem cells to regenerate human tissues [28,29]. It has been reported that MSC express markers pertaining to corneal epithelial stem cells [30-33] and can be induced to differentiate to epithelial like cells both *in vitro* and *in vivo* [33-36]. A critical factor in mesenchymal stem cell biology is to propagate cells with suitable stem cell characteristics, multipotency and high proliferative potential for its use in various biomedical applications. Under *in vitro* conditions, culture media plays a major role in growth, multiplication and maintenance of stemness of cells. Thus selection of optimal medium for culture and expansion of stem cells is crucial. In this study MSC were isolated and maintained

in different media formulations and cells cultured in a MEM showed more proliferation and maintenance potential of stem cells similar to previous reports [37]. Although no unique marker is known for MSC, their cell-surface antigen profile has been well explored. According to previous reports, MSC express markers such as ABCG2 [38], CD105 [39], vimentin [40] and CD90 [41]. The expression of ABCG2 was found to be localized in the cell membrane as well as in cytoplasm, as reported earlier [42]. Characteristic cytoskeletal expression of vimentin illustrated the filamentous nature and confirmed the significant role of vimentin in supporting and anchoring. It has been reported that great variation exists in the expression of CD90 which are reported to be

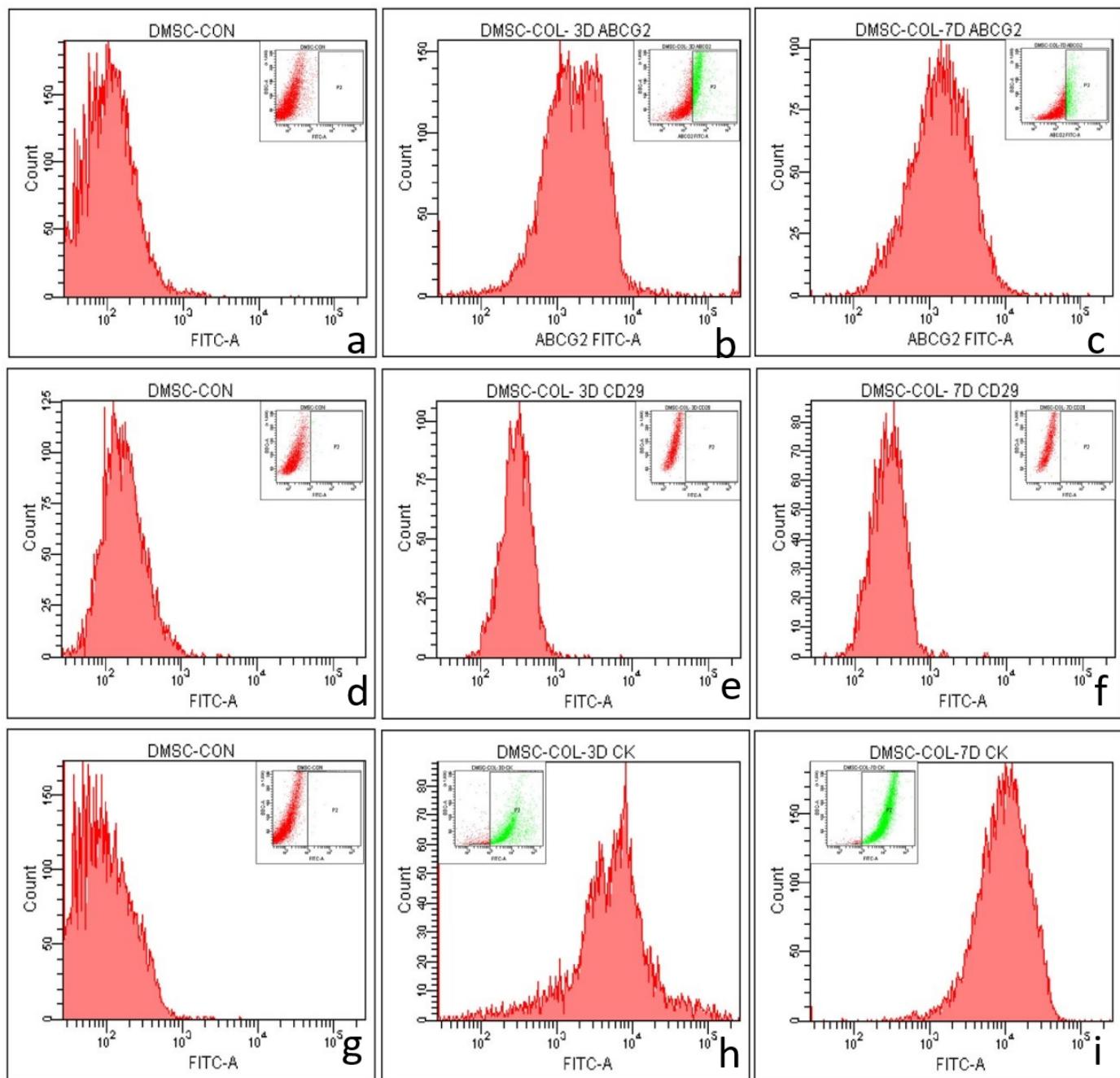


Figure 7. Flow cytometry analysis of MSC in CS_{cm} showing the expression of (a, b, c) ABCG2 (a) control (b) 3 day (c) 7 day, (d, e, f) CD29 (d) control (e) 3 day (f) 7 day and (g, h, i) CK3/12 (g) control (h) 3 day (i) 7 day. The inset image denotes the gated population.

expressed mostly by mesenchymal stromal cells [43]. The expression of CD90 was found to be negative, which we presume may be due to different stage of cell commitment and culturing conditions [37]. The immune reactivity obtained against ABCG2, CD105 and vimentin as well as the negative reaction for CD90 concord with previously reported findings [37].

Fibroblast like cells in the limbal stroma with self-renewal property and multipotency similar to MSC have been reported [44]. Here we describe the isolation and maintenance of rabbit LF with spindle shaped morphology and high proliferative potential. In order to determine the phenotype of isolated cells from limbal stroma of rabbits, the cells were characterized

by markers as reported elsewhere [8,37,43]. SPARC has been suggested as one of the markers expressed only by LF cells which have a functional role in maintenance of corneal homeostasis [43]. Immunocytochemical analysis of LF cultures showed the cytoplasmic, filamentous expression of vimentin suggesting its mesodermal origin. The expression of SPARC in the cytoplasm of cells confirmed that the isolated cells were LF. The results obtained in our study correlated well with previous reports [43]. However, negative expression of CD90 was noted for rabbit LF, which were contradictory to the findings of Ahmad *et al* in human fibroblasts [42]. The data obtained from our study suggest that the rabbit LF are negative for the expression of CD90 and hence can be regarded as a negative marker for the characterization of such cells in future.

Table 3. Expression levels of various antigens in different culture systems.

Culture System	CK3/12		ABCG2		CD29	
	3 rd Day	7 th Day	3 rd Day	7 th Day	3 rd Day	7 th Day
CS _{ecm}	0.5 ± 0.01%	0.2 ± 0.11%	97 ± 1.65%	98 ± 1.22%	53 ± 2.33%	53 ± 3.22%
CS _{ecm+cm}	86 ± 2.31%	86 ± 3.44%	34 ± 6.33%	39 ± 2.66%	1 ± 0.87%	1 ± 0.44%
CS _{cm}	84 ± 3.21%	88 ± 4.11%	36 ± 2.33%	21 ± 3.22%	0.3 ± 0.11%	0.4 ± 0.22%

Percentage of cells positive for CK3/12, ABCG2 and CD29 in culture systems (CS_{ecm}, CS_{ecm+cm}, CS_{cm}) revealed by flow cytometry analysis.

The stem cell maintenance and function are regulated by various intrinsic and extrinsic factors provided by a unique local microenvironment or niche. The limbal stem cells and their progeny that reside within small clusters in the basal epithelium in close spatial relationship with specific basement membrane and matrix components [44-48] as well as with stromal fibroblasts provides increased levels of growth and survival factors [41,49]. Previous reports demonstrate that it is possible to differentiate stem cells to corneal epithelial cells by simulating the limbal niche either by co culture, or in the presence of collagen and limbal fibroblast CM [50]. In order to replicate such culture conditions, *in vitro* transwell culture systems have been implemented to make use of secretory factors to aid in differentiation [51]. In our study, 3 different culture systems were formulated based on 2 parameters - ECM and conditioned medium. The corneal matrix is known to contain collagen-I [52] [53] and hence was used as one of the parameters. Expression levels of CK3/12 were used for evaluating differentiation into corneal epithelial phenotype. Results showed that collagen-I alone had no role in regulating cell differentiation, but favored the maintenance of stem cells as indicated by increased expression of ABCG2 and CD29. In CS_{ecm+cm}, the number of cells expressing CK3/12 was found to be increased with a concomitant reduction in the expression of stem cell markers. The CS_{cm} alone was also found to aid in differentiation and expression of corneal epithelial phenotype. We presume that the released factors produced by limbal stromal cells might have promoted MSC to differentiate into the corneal epithelium *in vitro*. The current findings shows that secretory factors by LF can induce transdifferentiation of bone marrow derived stem cells to corneal epithelial-like cells, as indicated by CK12 expression. The precise molecular mechanisms of differentiated reprogramming remains to be determined and further substantiation are required in *in vivo* models of LSCD. This study demonstrated that rabbit MSC could be induced into corneal epithelial-like cells *in vitro*, providing a new source of cells for the treatment of corneal disorders.

Conclusion

Mesenchymal stem cells could be transdifferentiated into corneal epithelial-like cells *in vitro* by simulating corneo-limbal niche. Transdifferentiating mesenchymal stem cells to corneal epithelial like cells will provide a useful, autologous source of adult stem cells in therapeutic approaches for patients suffering from bilateral limbal stem cell deficiency.

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