

Ambroxol hydrochloride, a chaperone therapy for Paget's disease of bone and other common autophagy-mediated aging diseases?

Mohamed Numan*

Centre Hospitalier Universitaire de Québec, Research Centre, Québec, QC, G1V 4G2, Canada

Introduction

Paget's disease of bone (PDB) with or without frontotemporal dementia shares common pathogenic mechanisms with some neurodegenerative diseases such as Parkinson's disease, amyotrophic lateral sclerosis (ALS), Alzheimer's, disease, and inclusion body myopathy [1]. These disorders are known to be related to autophagy machinery impairment [1]. Recent studies have reported roles of Ambroxol hydrochloride in treatment of some oxidative stress induced and autophagy mediated diseases such as Parkinson's disease (PD), related synucleinopathies, Dementia, Alzheimer's, Huntington's, Creutzfeldt-Jakob/prion, and Gaucher (GD) diseases [2-8].

The hypothesis

We made the hypothesis that Ambroxol hydrochloride should be a therapeutic avenue for PDB.

Evaluation of the hypothesis and discussion

Why Autophagy is essential for the normal cell biology?

Autophagy is essential process for normal cell biology. It is responsible for eliminating non-functional or misfolded proteins [1,8,9], and plays an important role in cell proliferation and cell death regulation [1, 8-10]; moreover it provides the energy needed for the cell and protect it against damaging by oxidative stress or starvation [1,11,12]. Autophagy can be initiated by some inducers such as excessive autophagic vacuoles, excessive aggregation of misfolded or unfolded proteins, accumulation of non-functional organelles, bacteria, virus, and by rapamycin (in vitro) [10,13]. The four main steps of autophagy are stimulation and initiation, nucleation, elongation, following by maturation and fusion between autophagosome and lysosome vacuoles initiators forming autolysosomes [1,10]. Autophagy machinery deficiency result in failure of elimination of misfolded/unfolded protein aggregates. Accumulation of non-functional misfolded proteins aggregates leads to symptoms of PDB, PD, related synucleinopathies, and other autophagy mediated aging diseases [1].

Mechanism of Autophagy impairment in PDB and in some autophagy mediated neurodegenerative disease:

SQSTM1/p62 gene mutations have been reported in some cases of ALS, PDB, and frontotemporal dementia [1,13-16]. Depriving the cell from full functional SQSTM1/p62 protein may lead to failing in autophagy functions; its role in eliminating misfolded/unfolded proteins aggregates and non-functional organelles [1,17]. In normal cells, Ubiquitin protein marks the non-functional proteins leading

to its elimination by autophagosome. SQSTM1/p62 is able to bind ubiquitinated proteins. It induces the autophagic elimination of ubiquitinated proteins by exporting them from the nucleus into the lysosomes [18]. Moreover, in nucleus, SQSTM1/p62 protein stimulates proteasomes to eliminate nuclear polyubiquitinated protein aggregates [1,17]. On the other hand, SQSTM1/p62 has a protective effect on huntingtin-induced cell death. Dysfunctional mutant Huntingtin protein and impairment of autophagy machinery may lead to Huntington's disease [19].

Role of autophagy impairment in PD:

Formation of Lewy's bodies (intraneuronal inclusions mainly consists of α -synuclein protein) leads to Parkinson's disease [1,19,20]. Glucocerebrosidase (GBA) gene mutations are considered the most risk factor that predisposes to PD [3]. Furthermore, some familial forms of Parkinson's disease caused by point mutations and multiplications of the whole locus at α -synuclein gene [21]. α -synuclein overexpression suppresses macroautophagy in mammalian cells and in transgenic mice leading to mitochondrial dysfunction, induces cell susceptibility to proapoptotic assaults, accumulate p62 protein, and increase proteins aggregation [22]. Moreover the excessive secretion of α -synuclein protein may leads to suppression of Rab1a protein function, impairment of 26S proteasome function [22]. Other studies reported its role in inhibition of ubiquitin-proteasome activity in vitro [22-25]. Impairment of autophagy pathway suppress dopamine secretion, which is the main cause of PD [22]. Taken together, we can conclude that mutant α -synuclein protein is highly involved in autophagy impairment.

Role of Ambroxol Hydrochloride

Recent studies reported important roles of Ambroxol Hydrochloride in GD and PD with GBA mutations. It increases the lysosomal enzyme mass, reduces dihydroethidium oxidation stress rate, improves of glucosylceramidase activity in fibroblasts with GBA mutation. Moreover, it suppresses α -synuclein overexpression neuroblastoma cell lines [2]. Furthermore, it is suggested to be one of chaperone therapies for protein misfolding diseases such as Gaucher disease [4], and for impairment of autophagy-lysosome system occurred in many

Correspondence to: Mohamed Numan, Centre Hospitalier Universitaire de Québec, Research Centre, Québec, QC, G1V 4G2, Canada, E-mail: Mohamednuman.ul@gmail.com; Mohamed.numan.1@ulaval.ca

Received: April 29, 2017; **Accepted:** May 25, 2017; **Published:** May 29, 2017

autophagy-mediated diseases [2]. Interestingly to mention that it's commercially available and widely used as a safe expectorant drug [4].

Consequently, we suggest a promised role for Ambroxol Hydrochloride in improving the autophagy process functions, decrease oxidative stress rate, and decrease the excessive osteoclasts activity, which is the main cause of sporadic and familial PDB.

References

1. Numan MS, Brown JP, Michou L (2015) Impact of air pollutants on oxidative stress in common autophagy-mediated aging diseases. *Int J Environ Res Public Health* 12: 2289-2305. [\[Crossref\]](#)
2. McNeill A, Magalhaes J, Shen C, Chau KY, Hughes D, et al. (2014) Ambroxol improves lysosomal biochemistry in glucocerebrosidase mutation-linked Parkinson disease cells. *Brain* 137: 1481-1495. [\[Crossref\]](#)
3. Schapira AH (2015) Glucocerebrosidase and Parkinson disease: Recent advances. *Mol Cell Neurosci* 66: 37-42. [\[Crossref\]](#)
4. Suzuki Y (2014) Emerging novel concept of chaperone therapies for protein misfolding diseases. *Proc Jpn Acad Ser B Phys Biol Sci* 90: 145-162. [\[Crossref\]](#)
5. Sybertz E, Krainc D (2014) Development of targeted therapies for Parkinson's disease and related synucleinopathies. *J Lipid Res* 55: 1996-2003. [\[Crossref\]](#)
6. Schapira AH, Gegg ME (2013) Glucocerebrosidase in the pathogenesis and treatment of Parkinson disease. *Proc Natl Acad Sci USA* 110: 3214-3215. [\[Crossref\]](#)
7. Maegawa GH, Tropak MB, Buttner JD, Rigat BA, Fuller M, et al. (2009) Identification and characterization of ambroxol as an enzyme enhancement agent for Gaucher disease. *J Biol Chem* 284: 23502-23516. [\[Crossref\]](#)
8. Glick D, Barth S, Macleod KF (2010) Autophagy: cellular and molecular mechanisms. *J Pathol* 221: 3-12. [\[Crossref\]](#)
9. Numan MS, Amiable N, Brown JP, Michou L (2015) Paget's disease of bone: an osteoimmunological disorder? *Drug Des Devel Ther* 9: 4695-4707. [\[Crossref\]](#)
10. Choi AM, Ryter SW, Levine B (2013) Autophagy in human health and disease. *N Engl J Med* 368: 1845-1846. [\[Crossref\]](#)
11. Karantza-Wadsworth V, Patel S, Kravchuk O, Chen G, Mathew R, et al. (2017) Autophagy mitigates metabolic stress and genome damage in mammary tumorigenesis. *Genes Dev* 21: 1621-1635. [\[Crossref\]](#)
12. Martindale JL, Holbrook NJ (2002) Cellular response to oxidative stress: Signaling for suicide and survival*. *J Cell Physiol* 192: 1-15. [\[Crossref\]](#)
13. Nikolettou V, Papandreou ME, Tavernarakis N (2015) Autophagy in the physiology and pathology of the central nervous system. *Cell Death Differ* 22: 398-407. [\[Crossref\]](#)
14. Le Ber I, Camuzat A, Guerreiro R, Bouya-Ahmed K, Bras J, et al. (2013) SQSTM1 mutations in French patients with frontotemporal dementia or frontotemporal dementia with amyotrophic lateral sclerosis. *JAMA Neurol* 70: 1403-1410. [\[Crossref\]](#)
15. Rubino E, Rainero I, Chiò A, Rogaeva E, Galimberti D, et al. (2012) SQSTM1 mutations in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Neurology* 79: 1556-1562. [\[Crossref\]](#)
16. Chung PYJ, Van Hul W (2012) Paget's disease of bone: evidence for complex pathogenetic interactions. *Semin Arthritis Rheum* 41: 619-641. [\[Crossref\]](#)
17. Lippai M, LÅ'w P (2014) The role of the selective adaptor p62 and ubiquitin-like proteins in autophagy. *Biomed Res Int* 2014: 832704. [\[Crossref\]](#)
18. Komatsu M, Ichimura Y (2010) Physiological significance of selective degradation of p62 by autophagy. *FEBS Lett* 584: 1374-1378. [\[Crossref\]](#)
19. Bjorkoy G, Lamark T, Brech A, Outzen H, Perander M, et al. (2005) p62/SQSTM1 forms protein aggregates degraded by autophagy and has a protective effect on huntingtin-induced cell death. *J Cell Biol* 171: 603-614. [\[Crossref\]](#)
20. Lynch-Day MA, Mao K, Wang K, Zhao M, Klionsky DJ (2012) The role of autophagy in Parkinson's disease. *Cold Spring Harb Perspect Med* 2: a009357. [\[Crossref\]](#)
21. Klein C, Schlossmacher MG (2006) The genetics of Parkinson disease: Implications for neurological care. *Nat Clin Pract Neurol* 2: 136-146. [\[Crossref\]](#)
22. Winslow AR, Chen CW, Corrochano S, Acevedo-Arozena A, Gordon DE, et al. (2010) alpha-Synuclein impairs macroautophagy: implications for Parkinson's disease. *J Cell Biol* 190: 1023-1037. [\[Crossref\]](#)
23. McNaught KS, Olanow CW, Halliwell B, Isacson O, Jenner P (2001) Failure of the ubiquitin-proteasome system in Parkinson's disease. *Nat Rev Neurosci* 2: 589-594. [\[Crossref\]](#)
24. McNaught KS, Belizaire R, Isacson O, Jenner P, Olanow CW (2003) Altered proteasomal function in sporadic Parkinson's disease. *Exp Neurol* 179: 38-46. [\[Crossref\]](#)
25. Snyder H, Mensah K, Theisler C, Lee J, Matouschek A, et al. (2003) Aggregated and monomeric alpha-synuclein bind to the S6' proteasomal protein and inhibit proteasomal function. *J Biol Chem* 278: 11753-11759. [\[Crossref\]](#)