Monitoring Regulatory T-Cells after Transplantation: Is It Useful?

Kathryn Brown and Wilson Wong

MRC Centre for Transplantation, King's College London School of Medicine at Guy's, King's and St Thomas' Hospitals, London, UK

Abstract

Currently, the gold standard for confirmation of rejection episodes in transplanted organs is histologic analysis of biopsy samples. There is no method to evaluate the risk of rejection in patients or to identify those who might be weaned off immunosuppression entirely. The development of biomarkers for use in transplantation would help achieve this goal of tailored immunosuppressive treatment for individual patients. Markers of regulatory T-cells, with their role in regulating the alloimmune response, have been investigated for their usefulness in this situation. This review will discuss the studies to date on the diagnostic and prognostic potential of regulatory T-cells in organ transplantation. (Trends in Transplant. 2009;3:119-28)

Corresponding author: Wilson Wong, wilson.wong@kcl.ac.uk

Key words

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Introduction

Surveillance of solid organ transplants, such as kidney and liver, for rejection and evaluation of graft function is routinely performed through functional parameters such as plasma creatinine and liver enzymes, respectively. Any deterioration in function in the absence of any other obvious causes usually prompts a biopsy to exclude rejection, especially within the first year of transplantation when the risk is high. This process has

Correspondence to:
Wilson Wong
MRC Centre for Transplantation
5th Floor, Tower Wing
Great Maze Pond
SE1 9RT London, UK
E-mail: wilson.wong@kcl.ac.uk

several disadvantages. Firstly, the biopsy procedure carries risks of complications, most seriously bleeding from the biopsy site, which may lead to graft loss or even death. Secondly, sampling error may result in inaccurate representation of the organ graft as a whole. Thirdly, by the time the transplanted organ is functionally compromised, irreversible tissue damage may have already taken place. The ideal method for monitoring rejection would be noninvasive and could detect rejection early. In the absence of such a monitoring method, histologic examination of biopsy samples remains the gold standard. It would be ideal to develop a more sophisticated screening method for rejection so that unnecessary biopsies could be avoided. In addition, it would be useful to establish the immunologic risks of individual patients after organ transplantation so that immunosuppressive therapy could be tailored accordingly. As

rejection is primarily an immune-mediated response, measurement of immunologic parameters has the potential to be developed to serve these purposes.

Biomarkers and immunologic risk

A large amount of effort has been devoted to the search for biomarkers (defined as a biological molecule whose presence can indicate a disease state or a high likelihood/risk of developing a disease or a specific disease phenotype¹) in many disease states and the field of transplantation is no exception. Initial studies concentrated on analyzing a few candidate genes such as the mediators of cytotoxic T-cell killing perforin and granzyme B^{2,3}. More recently, with advances in technology, there has been an increase in microarray based gene expression profiling, which examines the whole genome and does not require the identification of a possible target⁴.

The identification of one biomarker, or more likely a panel of biomarkers, to detect a rejection episode would not only allow earlier diagnosis and therefore intervention, but may also identify patients with different levels of risk by providing information on the antidonor response of the recipient.

A percentage of patients who discontinue their immunosuppression do not reject their grafts. This phenomenon is more common in liver transplant patients⁵⁻⁷, but also occurs in recipients of kidney transplants, although much less frequently^{8,9}. Much work has gone into defining a "tolerogenic profile", and this may allow not only the use of lower doses of immunosuppression in patients with tolerogenic profiles, but possibly the weaning of these patients off immunosuppression entirely. In contrast, patients at an increased risk of rejection according to the biomarker profile could be maintained on more intense immunosuppressive regimes to prevent rejection.

Currently, there is no way of predicting what level of immunosuppression a patient will need, which inevitably leads to some patients being over-immunosuppressed while others are under-immunosuppressed. The development of biomarkers is the first step to providing tailored immunosuppressive therapy for individual patients.

Regulatory T-cells

The importance of regulatory T-cells (T_{reg}) in tolerance models makes their surface or intracellular markers obvious candidates to provide a biomarker for transplantation. The characteristics of these cells and their role in the induction and maintenance of tolerance have been discussed elsewhere 10-12. Although the vast majority of organ recipients are not truly tolerant of their grafts, T_{req} may still play an important role in the prevention of rejection, adjuvant to the immunosuppressive drugs used in the clinic. If so, monitoring their activity in transplant recipients may provide useful immunologic information. This review will focus on their diagnostic potential, both in terms of aiding the diagnosis of rejection and providing an indication of prognosis and information to aid tailoring immunosuppressive therapy for individual patients.

To monitor T_{reg} , a robust marker should ideally be available. Unfortunately, a definitive marker for T_{reg} in humans is yet to be found. The classic T_{reg} are enriched within the CD4+CD25+ T-cell population^{13,14}. To date, the most reliable marker available is forkhead box p3 (FOXP3), a member of the forkhead/winged helix family of transcription factors¹⁵⁻¹⁷. In mice, Foxp3 is a specific marker of T_{reg} ¹⁵⁻¹⁷, however in humans this transcription factor is also transiently expressed in non-regulatory T-cells upon activation¹⁸. Other markers used to identify T_{reg} include CD45RB^{19,20}, CTLA-4^{21,22}, GITR²³⁻²⁵, CD122²⁶, CD103²⁷, and galectin-10²⁸, and the absence

Table 1. Summary of the use of FOXP3 mRNA measurement in human solid organ transplant recipients during rejection episodes to aid diagnosis or as a predictor of outcome

Author	Organ	Sample source (factor measured)	No. of patients with rejection/total	Diagnostic of rejection?	Predictive of better/ worse outcome?
Aquino-Dias, et al.31	Kidney	Intragraft (mRNA)	20/75	Yes	
		Blood (mRNA)	20/75	Yes	
		Urine (mRNA	20/75	Yes	
Ashton-Chess, et al.61	Kidney	Intragraft (mRNA)	14/48	No	
		Blood (mRNA)	15/205	No	
Grimbert, et al.32	Kidney	Intragraft (mRNA)	15 BL/11 AR/36 total	Yes	
Mansour, et al.47	Kidney	Intragraft (mRNA)	21/46		Better
Bestard, et al.48	Kidney	Intragraft (cell no.)	37		Better
Bunnag, et al.45	Kidney	Intragraft (mRNA)	31/95	Yes	No
Veronese, et al.44	Kidney	Intragraft (cell no.)	41/73	Yes	Worse
Martin, et al.46	Kidney	Intragraft (cell no.)	17/17	N/A	Better
Muthukumar, et al.51	Kidney	Urine (mRNA)	36/83	Yes	Better
Demirkiran, et al.33	Liver	Intragraft (mRNA)	3/20	Yes	
Sakamoto, et al.49	Liver	Blood (mRNA)	4/15		Better
Dijke, et al.34	Heart	Intragraft (mRNA)	26/41	Yes	
	Heart	Blood (mRNA)	26/41	No	

of CD127 can also indicate a regulatory phenotype 29,30 . A combination of these markers may be the best way of identifying $T_{\rm ren}$.

The use of FOXP3 as a biomarker has been investigated in several aspects of transplantation: the diagnosis of rejection; the prediction of rejection and the outcome of a rejection episode; the presence of tolerance; and involvement in graft-versus-host disease (GVHD).

Using regulatory T-cells to diagnose rejection

Given their potential to regulate allore-active T-cells, the presence of $T_{\rm reg}$ in higher numbers should, in theory, exert a graft-protective effect. Conversely, low numbers of $T_{\rm reg}$ may suggest rejection.

Intragraft T_{reg}/FOXP3

Results from these studies are summarized in table 1. Aquino-Dias, et al. studied kidney allografts that had suffered from delayed graft function. The levels of FOXP3 mRNA were paradoxically significantly higher in those patients undergoing acute rejection, and indeed were a more reliable marker of rejection than either of the cytotoxic mediators perforin and granzyme B³¹. A separate comparison of kidney transplant recipients with acute rejection and borderline changes found that again FOXP3 mRNA levels were higher in the rejection group³².

High FOXP3 mRNA levels have also been found to be predictive of rejection in liver allografts³³. However, FOXP3 levels were also elevated during hepatitis C virus infection, suggesting that FOXP3 would not provide the

level of specificity required. Levels of FOXP3 mRNA have also been examined in heart transplant recipients³⁴. Again, higher FOXP3 mRNA levels were present in the 26 acutely rejecting patients, compared to the 15 patients without rejection.

The increase in $T_{\rm reg}$ in rejection probably reflects an overall increase in the immune response, with $T_{\rm reg}$ being a portion of this, rather than acting in its graft-protective capacity.

There is now increasing evidence that intragraft mRNA levels of FOXP3 do seem to be a good indicator of acute rejection. However, to measure intragraft mRNA, biopsies must still be performed and, so, rejection can be diagnosed by histologic examination, which is still regarded as the gold standard. Therefore, in this respect, it serves little clinical purpose but is likely to remain as a useful research tool. Measuring FOXP3 mRNA levels in peripheral blood or urine (in the case of kidney transplantation) may circumvent this problem.

Peripheral blood and urine T_{red} /FOXP3

Aquino-Dias, et al., as well as studying intragraft FOXP3 mRNA, measured peripheral blood and urine FOXP3 mRNA levels in their cohort of patients with delayed graft function. They found that, as with intragraft FOXP3, mRNA levels of FOXP3 in blood and urine were significantly higher in the group undergoing acute rejection³¹, with sensitivity and specificity in blood of 94 and 95%, respectively, while both corresponding figures in urine were 100%. Satoda, et al. made a similar finding in a miniature swine model of lung transplantation³⁵.

In contrast, no difference in the levels of FOXP3 mRNA were found in the blood of

heart transplant recipients, despite there being a difference in intragraft FOXP3³⁴. Similar findings were made in liver transplant recipients³³. This may be due to the egress of these cells out of the blood and into the graft.

Peripheral blood and urine markers of T_{reg} therefore appear to have potential to be used in the diagnosis of acute rejection of kidney, and potentially lung, transplants, although they have not proven useful so far in heart or liver transplantation. However, the number of patients in the study is small and needs to be confirmed by others. The sensitivity and specificity of 100% in urine observed in the study is very impressive; it is difficult to envisage any tests with such high accuracies. Expansion of the cohort size is likely to reduce these figures. Nonetheless, urinary FOXP3 mRNA level should prove to be a useful marker in this respect.

The development of this method would allow a quick, noninvasive screening test for rejection, and $T_{\rm reg}$ levels could be monitored alongside functional indicators such as plasma creatinine, allowing a patient's immunologic status to be measured over time, and correlated to graft function.

Stem cell transplantation

The possibility of using T_{reg} to diagnose GVHD in recipients of allogeneic stem cell transplants has also been explored. In contrast to the situation with acute rejection of solid organ transplants, FOXP3 mRNA levels were found to be significantly reduced in all forms (acute, chronic, allo, auto) of GVHD in two independent studies^{36,37}. These data were contradicted by Meignin, et al., who found no difference in blood mRNA FOXP3 levels between patients with or without GVHD³⁸. More work will need to be carried out to clarify the situation, although Rieger, et al. examined FOXP3+ cells in intestinal GVHD lesions and

found evidence to support the hypothesis that lower numbers of FOXP3+ cells (relative to CD8+ cells) is indicative of GVHD³⁹.

Other studies have been carried out to investigate T_{reg} in GVHD, but these have used CD25 (in combination with CD4) to identify T_{reg}^{40-43} . Again, results from these studies have been contradictory, suggesting that CD25, with its expression on activated T-cells, is not a specific enough marker of T_{reg} .

Regulatory T-cells as predictors of rejection/rejection outcome

Intragraft

The presence of T_{reg} in biopsies appears to indicate rejection, presumably by revealing the extent of the alloimmune response. It may be expected that, further to this, higher levels of FOXP3 may indicate a better-regulated immune response and therefore predict a better outcome of the rejection episode. Data acquired so far is summarized in table 1.

This theory was contradicted by data acquired by Veronese, et al., who found by immunohistochemistry that, as expected, the 41 kidney allografts with acute cellular rejection had significantly more FOXP3+ cells than the 32 patients with acute humoral rejection⁴⁴. However, in the group with acute cellular rejection, the higher the number of FOXP3+ cells the worse the outcome. The numbers of CD4+ and CD8+ cells were also higher, suggesting that the higher number of FOXP3+ cells represented merely a more vigorous alloimmune response rather than a disproportionate number of FOXP3+ cells. They also found FOXP3+ T-cells infiltrating tubules, and coined the phrase T_{reg} tubulitis⁴⁴. Indeed, they found that FOXP3+ cells were more likely to infiltrate tubules than CD4+FOXP3- cells. This infiltration of tubules provides a mechanism for the

entry of T_{reg} into the urine (see next section). A separate independent study on FOXP3 mRNA in renal biopsies also found that FOXP3 mRNA levels did not correlate with good graft outcome⁴⁵.

Contrasting data came from a much smaller study by Martin, et al., who showed that of 17 kidney transplant biopsies studied, three underwent rejection resulting in graft loss within the first year⁴⁶. These three samples had no FOXP3+ cells when biopsied during the early stages of acute rejection. This study, although small, suggests that it may be worthwhile to conduct further investigations.

Levels of FOXP3 may be able to predict future rejection episodes in certain patients. A study of renal allografts with borderline changes determined that those grafts which did not progress to rejection had significantly higher levels of FOXP3 mRNA than those grafts which did progress⁴⁷. Therefore, in this particular group of patients with borderline changes, FOXP3 may be indicative of the likelihood of progression to rejection. Similarly, in a study of 37 cases of kidney biopsies with subclinical rejection, higher numbers of FOXP3+ predicted better graft function⁴⁸. FOXP3 may be useful in defining the immunologic status in these patients, which are currently not well understood, and predicting the chances of a rejection episode so that immunosuppression can be altered accordingly.

Peripheral blood

A time-course study was carried out on levels of FOXP3 mRNA in blood from liver transplant recipients. In all 15 patients, FOXP3 mRNA levels increased by day 7 posttransplantation, and then returned to baseline by day 28. In the four patients who went on to develop T-cell mediated rejection within 60 days, FOXP3 mRNA levels returned to baseline by day 14 posttransplantation⁴⁹. This study was

small and did not show any statistically significant differences. However, it does suggest that FOXP3, as part of a large panel of biomarkers, may be useful in the prediction of rejection.

Peripheral blood from heart transplant recipients pretransplantation was collected and $T_{\rm reg}$ function examined. The CD4+CD25hi cells from blood samples taken pretransplantation from heart transplant recipients who went on to develop acute rejection were less suppressive than those from non-rejecters⁵⁰. However, this does limit the usefulness of $T_{\rm reg}$ in blood to predict heart rejection as a functional assay of $T_{\rm reg}$ is probably too time-consuming to be performed clinically.

Urine

Muthukumar, et al. conducted a study into FOXP3 mRNA in the urine of kidney transplant recipients⁵¹. Again confirming the ability of FOXP3 mRNA levels to diagnose rejection, FOXP3 mRNA was higher in patients with rejection than those without and controls. Increased levels of FOXP3 mRNA within the urine of the rejection group was associated with better graft outcome. It is possible that the difference between the Veronese, et al. and Muthukumar, et al. data is due to the fact that the urinary mRNA levels represent only those T_{reg} that infiltrate tubules within the kidney.

Stem cell transplantation

To predict GVHD after stem cell transplantation, donor cell infiltrates, rather than the peripheral blood of recipients, were analyzed for FOXP3. Two independent studies on donor infiltrate in HLA-identical stem cell transplants showed that patients who received infiltrate with low numbers of CD4+FOXP3+ cells were more likely to suffer from severe GVHD^{52,53}.

A separate study used CD4 and CD25 to define T_{reg} , and found, conversely, that GVHD was associated with high numbers of CD4+CD25+ cells within the donor infiltrate⁵⁴. However, this again may be due to the use of CD25 as a marker of T_{reg} , as it was shown by Rezvani et al. that numbers of CD4+FOXP3+ and CD4+CD25+ cells in donor cell infiltrates were inversely correlated⁵².

The ability of FOXP3 to predict graft outcome in those patients undergoing rejection remains unclear. It does appear though that FOXP3 has greater potential as a predictor of future rejection episodes, both in kidney and liver transplant recipients, and may help to more successfully diagnose those patients with only slight histologic changes. FOXP3 within the donor infiltrate may also be useful in the prediction of GVHD.

Regulatory T-cells as biomarker of tolerance

As discussed earlier, some patients who discontinue their immunosuppression do not reject their grafts. This is known as spontaneous operational tolerance (SPOT), and great effort has gone into attempts to characterize these patients and define a tolerogenic profile. The results to date are summarized in table 2. This is a difficult task, given the rarity of these patients and the problem of obtaining samples from them.

Intragraft

Sachs, et al. have used a non-myeloablative conditioning regime prior to kidney and bone marrow transplantation, which has allowed them to discontinue immunosuppression at about one year post transplantation⁵⁵. Grafts continued to function well until studied here at 2-5.3 years posttransplantation. Levels of FOXP3 mRNA in kidney biopsies were

Author	Organ	Sample source	No. of SPOT patients/total	Marker of tolerance?
Kawai, et al.55	Kidney	Intragraft (mRNA)	6/14	Yes
Louis, et al.59	Kidney	Blood (cell no. and mRNA)	8/65	No
Alvarez, et al.67	Kidney	Blood (mRNA)	3/40	No
Brouard, et al.60	Kidney	Blood (mRNA)	17/75	No
Li, et al. ⁵⁸	Liver	Intragraft (mRNA)	28/64	No
		Intragraft (cell no.)	28/64	Yes
Pons, et al.64	Liver	Blood (cell no. and mRNA)	5/12	Yes

about six-times higher in the immunosuppression-free group than in the group with good graft function that were still receiving immunosuppressive therapy. This indicates that in the absence of immunosuppression, bone marrow transplantation can induce a regulatory phenotype. However, the number of patients is small and needs to be extended to confirm this important finding.

Given the ethical considerations of performing kidney biopsies on patients with good graft function, we⁵⁶ and others⁵⁷ have used a murine kidney transplant model, in which DBA/2 kidneys are spontaneously accepted by C57BL/6 recipients. High numbers of Foxp3+ cells were found in the graft and spleen of these mice. Although allografts were tolerated initially, some recipients underwent chronic rejection resulting in graft loss, and kidney allografts from these mice contained low numbers of Foxp3+ cells⁵⁶.

In liver transplant recipients, FOXP3 mRNA levels in biopsies were higher in grafts from SPOT patients than in those being maintained on immunosuppression but similar to those in chronically rejected grafts⁵⁸. However, when the FOXP3 was examined at the protein level, the number of FOXP3+ cells was significantly higher in SPOT patients than in both those on immunosuppression and those that had been chronically rejected. The reason

for this discrepancy is unknown as other studies have found good correlation between mRNA and cell numbers.

Peripheral blood

Louis, et al. investigated CD4+CD25hi cell numbers and FOXP3 mRNA in SPOT kidney transplant patients⁵⁹. Numbers of CD4+CD25hi cells and FOXP3 mRNA levels were similar in SPOT patients and those on immunosuppression. However, cell numbers and mRNA levels were lower in chronically rejecting patients. Similar results were obtained in a separate study⁵⁹, and Brouard, et al. reached the same conclusion using peripheral blood gene expression profiles⁶⁰. This ability to distinguish chronic rejection using T_{reg} has been contradicted, however, by a study by Ashton-Chess, et al.⁶¹.

A Europe-wide study also analyzed mRNA levels in SPOT kidney transplant recipients⁶². They found that although FOXP3 mRNA levels alone could not be used to distinguish SPOT patients, this group did have a significantly higher ratio of FOXP3 to alphamannosidase mRNA than chronically rejecting patients and those with stable function on immunosuppression. This confirms the importance of examining several parameters when judging the immunologic profile of a patient.

Peripheral blood from liver transplant SPOT patients contained higher numbers of CD4+CD25+ cells than liver transplant recipients on immunosuppression⁶³. In addition, CD4+CD25hi cell numbers and FOXP3 mRNA levels increased and remained high upon withdrawal of immunosuppression in five liver transplant recipients who did not reject their grafts. In the seven patients who underwent rejection upon withdrawal, neither CD4+CD25hi cell numbers nor FOXP3 mRNA levels increased 64 . Therefore, T_{req} may be more indicative of tolerance in liver compared to kidney transplant recipients, and could possibly be used to identify the significant percentage of liver transplant recipients currently on immunosuppression who could be candidates for weaning (estimated to be up to 20%65), and to determine the success, or failure, of this process.

Conclusions

Although a great deal of conflicting data has been published on the use of FOXP3 as a biomarker in transplantation, it seems that using FOXP3 mRNA levels in peripheral blood could become a reliable method of screening for acute rejection of kidney grafts and GVHD after stem cell transplantation. Larger scale studies involving more patients from multiple centers are urgently needed to establish this. However, it is unlikely that monitoring of T_{rea} will completely replace histologic examination of the transplanted organs, but may provide additional information on the immune status of the recipients during rejection episodes. In those kidney transplant patients with only slight histologic abnormalities, FOXP3 analysis may help to clarify the status of the immune response and predict future rejection episodes. However, the potential of FOXP3 as a predictor of graft outcome is still unclear, given the number of small studies yielding conflicting results. Again, cooperation between different transplant centers to perform a large multicentre study would clarify this issue.

Some of the conflicting data described above may be related to other factors that affect FOXP3 expression. Two studies have shown that FOXP3 is increased with time after transplantation^{45,61}. Factors such as these must be considered before FOXP3 is used for clinical diagnostic purposes.

It is very unlikely that one single biomarker, no matter how important in the alloimmune response, will be able to specifically and sensitively predict rejection and graft outcome. A panel of biomarkers from different facets of the immune response is probably the way forward. Genes which have shown promise include those for perforin, granzyme B, and granulysin, which reflect cytotoxic T-cell activity; NKG2D, an activating receptor on natural killer cells and T-cells; cytokines such as interferon-y; and the chemokine IP-10 (reviewed⁶⁶). It is likely that FOXP3, or more definitive marker(s) of T_{req}, will be an important component of this panel. Much more work needs to be done in this area, but the ultimate goal is within our grasp. When achieved, the greatest potential for this technique would be the assessment of the individual immunologic risks of transplant recipients, thus enabling the tailoring of immunosuppression to maximize graft outcome while minimizing side effects.

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References

- Kurian S, Grigoryev Y, Head S, et al. Applying genomics to organ transplantation medicine in both discovery and validation of biomarkers. Int Immunopharmacol. 2007;7:1948-60.
- Sharma VK, Bologa RM, Li B, et al. Molecular executors of cell death--differential intrarenal expression of Fas ligand, Fas, granzyme B, and perforin during acute and/or chronic rejection of human renal allografts. Transplantation. 1996;62:1860-6.
- 3. Li B, Hartono C, Ding R, et al. Noninvasive diagnosis of renal-allograft rejection by measurement of messenger RNA

- for perforin and granzyme B in urine. N Engl J Med 2001:344:947-54.
- Sarwal M, Chua MS, Kambham N, et al. Molecular heterogeneity in acute renal allograft rejection identified by DNA microarray profiling. N Engl J Med. 2003;349:125-38.
- Ramos HC, Reyes J, Abu-Elmagd K, et al. Weaning of immunosuppression in long-term liver transplant recipients. Transplantation. 1995;59:212-17.
- Takatsuki M, Uemoto S, Inomata Y, et al. Weaning of immunosuppression in living donor liver transplant recipients. Transplantation. 2001;72:449-54.
- Mazariegos G, Reyes J, Marino I, et al. Weaning of immunosuppression in liver transplant recipients. Transplantation. 1997;63:243-9.
- Uehling DT, Hussey JL, Weinstein AB, Wank R, Bach FH. Cessation of immunosuppression after renal transplantation. Surgery. 1976;79:278-82.
- Zoller KM, Cho SI, Cohen JJ, Harrington JT. Cessation of immunosuppressive therapy after successful transplantation: a national survey. Kidney Int. 1980;18:110-14.
- Wood KJ, Sakaguchi S. Regulatory T cells in transplantation tolerance. Nat Rev Immunol. 2003;3:199-210.
- 11. Tang A, Bluestone JA. The Foxp3+ regulatory T cell: a jack of all trades, master of regulation. Nat Immunol. 2008;9:239-44.
- Long E, Wood KJ. Understanding FOXP3: progress towards achieving transplantation tolerance. Transplantation. 2007:84:459-61.
- Hall B, Pearce N, Gurley K, Dorsch S. Specific unresponsiveness in rats with prolonged cardiac allograft survival after treatment with cyclosporine. III. Further characterisation of the CD4* suppressor cell and its mechanism of action. J Exp Med. 1990;171:141-57.
- Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor a-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. J Immunol. 1995;155:1151-64.
- Hori S, Namura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. Science. 2003;299:1057-61.
- Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4*CD25* regulatory T cells. Nat Immunol. 2003;4:330-6.
- Khattri R, Cox T, Yasayko SA, Ramsdell F. An essential role for Scurfin in CD4*CD25* regulatory T cells. Nat Immunol. 2003;4:337-42.
- Morgan ME, van Bilsen JH, Bakker AM, et al. Expression of FOXP3 mRNA is not confined to CD4*CD25* T regulatory cells in humans. Hum Immunol. 2005;66:13-20.
- Morrissey PJ, Charrier K, Braddy S, Liggitt D, Watson JD. CD4* T cells that express high levels of CD45RB induce wasting disease when transferred into congenic severe combined immunodeficient mice. Disease development is prevented by cotransfer of purified CD4* T cells. J Exp Med. 1993;178:237-44.
- Powrie F, Leach MW, Mauze S, Caddle LB, Coffman RL. Phenotypically distinct subsets of CD4* T cells induce or protect from chronic intestinal inflammation in C. B-17 scid mice. Int Immunol. 1993;5:1461-71.
- Read S, Malmstrom V, Powrie F. Cytotoxic T lymphocyteassociated antigen 4 plays an essential role in the function of CD25+CD4+ regulatory cells that control intestinal inflammation. J Exp Med. 2000;192:295-302.
- Takahashi T, Tagami T, Yamazaki S, et al. Immunologic self-tolerance maintained by CD25(+)CD4(+) regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. J Exp Med. 2000;192:303-10.
- Gavin MA, Clarke SR, Negrou E, Gallegos A, Rudensky A. Homeostasis and anergy of CD4(+)CD25(+) suppressor T cells in vivo. Nat Immunol. 2002;3:33-41.
- McHugh RS, Whitters MJ, Piccirillo CA, et al. CD4(+)CD25(+) immunoregulatory T cells: gene expression analysis reveals a functional role for the glucocorticoid-induced TNF receptor. Immunity. 2002;16:311-23.
- Shimizu J, Yamazaki S, Takahashi T, Ishida Y, Sakaguchi S. Stimulation of CD25(+)CD4(+) regulatory T cells through GITR breaks immunological self-tolerance. Nat Immunol. 2002;3:135-42.

- Stephens LA, Mottet C, Mason D, Powrie F. Human CD4(+) CD25(+) thymocytes and peripheral T cells have immune suppressive activity in vitro. Eur J Immunol. 2001;31:1247-54.
- Suffia I, Reckling SK, Salay G, Belkaid Y. A role for CD103 in the retention of CD4+CD25+ Treg and control of leishmania major infection. J Immunol. 2005;174:5444-55.
- Kubach J, Lutter P, Bopp T, et al. Human CD4+CD25+ regulatory T cells: proteome analysis identifies galectin-10 as a novel marker essential for their anergy and suppressive function. Blood. 2007;110:1550-8.
- Liu W, Putnam AL, Xu-yu Z, et al. CD127 expression inversely correlates with FoxP3 and suppressive function of human CD4+ Treg cells. J Exp Med. 2006;203:1701-11.
- Seddiki N, Santner-Nanan B, Martinson J, et al. Expression of interleukin (IL)-2 and IL-7 receptors discriminates between human regulatory and activated T cells. J Exp Med. 2006;203:1693-700.
- 31. Aquino-Dias EC, Joelsons G, da Silva DM, et al. Non-invasive diagnosis of acute rejection in kidney transplants with delayed graft function. Kidney Int. 2008;73:877-84. *A systematic investigation of blood, urine and intragraft mRNA levels of various markers including FOXP3 in kidney transplant recipients showing that FOXP3 was the most accurate marker studied of rejection with high sensitivity and specificity.
- Grimbert P, Mansour H, Desvaux D, et al. The regulatory/ cytotoxic graft-infiltrating T cells differentiate renal allograft borderline change from acute rejection. Transplantation. 2007;83:341-6.
- Demirkiran A, Baan CC, Kok A, Metselaar HJ, Tilanus HW, van der Laan LJ. Intrahepatic detection of FOXP3 gene expression after liver transplantation using minimally invasive aspiration biopsy. Transplantation. 2007;83:819-23.
- 34. Dijke IE, Caliskan K, Korevaar SS, et al. FOXP3 mRNA expression analysis in the peripheral blood and allograft of heart transplant patients. Transplant Immunol. 2008;18:250-4.
- Satoda N, Shoji T, Wu Y, et al. Value of FOXP3 expression in peripheral blood as rejection marker after miniature swine lung transplantation. J Heart Lung Transplant. 2008;27:1293-301.
- 36. Miura Y, Thoburn CJ, Bright EC, et al. Association of Foxp3 regulatory gene expression with graft-versus-host disease. Blood. 2004;104:2187-93.
- Zorn E, Kim HT, Lee SJ, et al. Reduced frequency of FOXP3+ CD4+CD25+ regulatory T cells in patients with chronic graftversus-host disease. Blood. 2005;106:2903-11.
- 38. Meignin V, de Latour RP, Zuber J, et al. Numbers of Foxp3-expressing CD4+CD25high T cells do not correlate with the establishment of long-term tolerance after allogeneic stem cell transplantation. Exp Hematol. 2005;33):894-900.
- 39. Rieger K, Loddenkemper C, Maul J, et al. Mucosal FOXP3+ regulatory T cells are numerically deficient in acute and chronic GvHD. Blood. 2006;107:1717-23.
- Clark FJ, Gregg R, Piper K, et al. Chronic graft-versus-host disease is associated with increased numbers of peripheral blood CD4+CD25high regulatory T cells. Blood. 2004;103: 2410-16
- Sanchez J, Casano J, Alvarez MA, et al. Kinetic of regulatory CD25high and activated CD134⁺ (OX40) T lymphocytes during acute and chronic graft-versus-host disease after allogeneic bone marrow transplantation. Br J Haematol. 2004;126:697-703.
- 42. Schneider M, Munder M, Karakhanova S, Ho AD, Goerner M. The initial phase of graft-versus-host disease is associated with a decrease of CD4+CD25+ regulatory T cells in the peripheral blood of patients after allogeneic stem cell transplantation. Clin Lab Haematol. 2006;28:382-90.
- Nadal E, Garin M, Kaeda J, et al. Increased frequencies of CD4+CD25high Tregs correlate with disease relapse after allogeneic stem cell transplantation for chronic myeloid leukemia. Leukemia. 2007;21:472-9.
- 44. Veronese F, Rotman S, Smith RN, et al. Pathological and clinical correlates of FOXP3+ cells in renal allografts during acute rejection. Am J Transplant. 2007;7:914-22. **The authors detected the presence of FOXP3 positive cells infiltrating tubules and coined the phrase T_{reg} tubulitis. This forms the physical basis for the appearance of FOXP3 mRNA in the urine studied by several other investigators.

- Bunnag S, Allanach K, Jhangri GS, et al. FOXP3 expression in human kidney transplant biopsies is associated with rejection and time post transplant but not with favorable outcomes. Am J Transplant. 2008;8:1423-33.
- Martin L, de la Vega MF, Bocrie O, et al. Detection of Foxp3+ cells on biopsies of kidney transplants with early acute rejection. Transplant Proc. 2007;39:2586-8.
- Mansour H, Homs S, Desvaux D, et al. Intragraft levels of Foxp3 mRNA predict progression in renal transplants with borderline change. J Am Soc Nephrol. 2008;19:2277-81.
- Bestard O, Cruzado JM, Rama I, et al. Presence of FoxP3+ regulatory T cells predicts outcome of subclinical rejection of renal allografts. J Am Soc Nephrol. 2008; 19:2020-6.
- Sakamoto R, Asonuma K, Zeledon Ramirez ME, et al. Forkhead box P3 (FOXP3) mRNA expression immediately after living-donor liver transplant. Exp Clin Transplant. 2009;7:8-12.
- Dijke I E, Korevaar SS, Caliskan K, et al. Inadequate immune regulatory function of CD4+CD25bright+FoxP3+ T cells in heart transplant patients who experience acute cellular rejection. Transplantation. 2009;87:1191-200.
- 51. Muthukumar T, Dadhania D, Ding R, et al. Messenger RNA for FOXP3 in the urine of renal-allograft recipients. N Engl J Med. 2005;353:2342-51. **Using real time RT-PCR to measure the level of FOXP3 mRNA in urine of recipients of kidney transplant, the authors found that high levels were associated with better outcome.
- Rezvani K, Mielke S, Ahmadzadeh M, et al. High donor FOXP3-positive regulatory T-cell (Treg) content is associated with a low risk of GVHD following HLA-matched allogeneic SCT. Blood. 2006;108:1291-7.
- Wolf D, Wolf AM, Fong D, et al. Regulatory T-cells in the graft and the risk of acute graft-versus-host disease after allogeneic stem cell transplantation. Transplantation. 2007:83:1107-13
- 54. Stanzani M, Martins SLR, Saliba RM, et al. CD25 expression on donor CD4+ or CD8+ T cells is associated with an increased risk for graft-versus-host disease after HLA-identical stem cell transplantation in humans. Blood. 2004;103: 1140-6.

- Kawai T, Cosimi AB, Spitzer TR, et al. HLA-mismatched renal transplantation without maintenance immunosuppression. N Engl J Med. 2008;358:353-61.
- Brown K, Moxham V, Karegli J, et al. Ultra-localization of Foxp3+ T cells within renal allografts shows infiltration of tubules mimicking rejection. Am J Pathol. 2007;171:1915-22.
- Bickerstaff AA, Wang J-J, Pelletier RP, Orosz CG. Murine renal allografts: spontaneous acceptance is associated with regulated T cell-mediated immunity. J Immunol. 2001;167:4821-7.
- Li Y, Zhao X, Cheng D, et al. The presence of Foxp3 expressing T cells within grafts of tolerant human liver transplant recipients. Transplantation. 2008;86:1837-43.
- 59. Louis S, Braudeau C, Giral M, et al. Contrasting CD25^{hi}CD4⁺ T cells/FOXP3 patterns in chronic rejection and operational drug-free tolerance. Transplantation. 2006;81:398-407.
- Brouard S, Mansfield E, Braud C, et al. Identification of a peripheral blood transcriptional biomarker panel associated with operational renal allograft tolerance. Proc Nat Acad Sci USA. 2007;104:15448-53.
- Ashton-Chess J, Dugast E, Colvin RB, et al. Regulatory, effector, and cytotoxic T cell profiles in long-term kidney transplant patients. J Am Soc Nephrol. 2009;20:1113-22.
- Hernandez-Fuentes M, Sawitszki B, Sagoo P, et al. Indices of tolerance: interim report (abstract). Am J Transplant. 2006:6:403.
- Martinez-Llordella M, Puig-Pey I, Orlando G, et al. Multiparameter immune profiling of operational tolerance in liver transplantation. Am J Transplant. 2007;7:309-19.
- 64. Pons J A, Revilla-Nuin B, Baroja-Mazo A, et al. FoxP3 in peripheral blood is associated with operational tolerance in liver transplant patients during immunosuppression withdrawal. Transplantation. 2008;86:1370-8.
- 65. Lerut J, Sanchez-Fueyo A. An appraisal of tolerance in liver transplantation. Am J Transplant. 2006;6:1774-80.
- Anglicheau D, Suthanthiran M. Noninvasive prediction of organ graft rejection and outcome using gene expression patterns. Transplantation. 2008;86:192-9.
- Alvarez CM, Opelz G, Garcia LF, Susal C. Expression of regulatory T-cell-related molecule genes and clinical outcome in kidney transplant recipients. Transplantation. 2009;87:857-63.