First report of homozygous factor VII Padua (Arg304Gln) defect in a family from Argentina

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Abstract
Congenital FVII deficiency is the most frequently encountered defect among the Rare Bleeding Disorders (RBD). Geographical distribution is uneven but it has been described worldwide. Factor FVII Padua (Arg304Gln) is a Type 2 variant with shows FVII activity levels which varies with the origin of the thromboplastin used in the assay system. It is low (less than 5%) using rabbit brain thromboplastins, but normal (100%) with OX-brain preparations. Thromboplastins of human origin (placenta) or human recombinant reagents yield intermediate levels (40-50% of normal). FVII antigen is normal. FVII Padua has been described in many parts of the world but it seems rare in Latin America where only a few heterozygotes cases have been described but no homozygotes. We described here the first FVII Padua case seen in Argentina and, probably, in Latin America. The proposita is a 64 year old female who had a Spanish background and no bleeding tendency. She was found to be homozygote for the Arg304Gln mutation whereas her daughter was heterozygote and also asymptomatic. The reasons for the rarity of this mutation in Argentina and in Latin America are discussed.

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brother of the proposita (result of her father previous marriage) is reported to have frequent epistaxis but it is not known if the patient has a clotting defect.

There are no other cases of bleeding tendency in the family. The proposita’s husband died at the age of 19 because of drowning but he had a negative personal and family history for bleeding. The proposita underwent appendectomy in young age without undue bleeding; tooth extractions were also referred not to be accompanied by bleeding. Menstruations were normal and delivery by cesarean section at the age of 20 was uneventful. A diagnosis of FVII deficiency was made at the age of 60 in preparation for a colonoscopy. Because of this diagnosis, the patient underwent video colonoscopies under the protection of two aFVII concentrates (dosage unknown) without any undue bleeding or side effects. During this procedure a small polyp was removed. The control procedure was carried out 2 years later, under the protection of a total doses of 1.3 mg of aFVII concentrate. Again there was no bleeding or side effects and the procedure failed to show relapse of polyps.

At present the patient is in good health and has no bleeding manifestations.

The daughter of the proposita is 44 year old and has never presented with any bleeding symptom. Her menstrual cycles are also reported as normal.

There are no known episodes of venous or arterial thrombosis in the patients or in the family. aPTT and PT were carried out using standard procedures. The reagent used for the aPTT was supplied by Instrumentation Laboratory, Milan, Italy. Seven reagents were used for the PT and FVII assay, namely: three rabbit brain thromboplastins (STA Neoplastin plus, Stago Laboratories, Asniers, France; PT- Fibrinogen HS Plus, Instrumentation Laboratory, Milan, Italy; Tromboplastina S, hemo medica, Fisher Diagnostics, USA); a reagent obtained from human placenta (Thromborel S, Dade-Behring, Marburg, Germany) and two recombinant human thromboplastins (Dade-Innovin, Siemens, Marburg, Germany; and Recombiplastin 2G, Instrumentation Laboratory, Milan, Italy). An ox-brain thromboplastin was also used (Thrombotest, Nygaard Laboratories, Oslo, Norway).

Factor VII clotting assay were carried out on 1:10 diluted plasma using known FVII deficient plasma as substrate and the different thromboplastins. FVII antigens level was studied by an Elisa method (Asserachrom FVII, Stago Laboratories, Asniers, France).

DNA was extracted from dried thick drops of whole blood blotted on Whatman paper. For this purpose we used the kit (QiAMP DNA minikit) supplied by QIAGEN Laboratories (Qiagen s.r.l., Milan, Italy).

Amplification of exons 1 to 8 and respective splice junctions of the FVII gene were performed using oligonucleotide primers kindly supplied by Dr. James H. (Tyler, Tx, U.S.A.) or acquired from Invitrogen (Carlsbad, Ca, U.S.A.).

Mutational analysis was performed by polymerase chain reaction (PCR) amplification using oligo 8AF (5’-GAGGTGGCAAGGTGTTGGAAAA-3’), 8AR (5’CGGCAACACAGATCGTACTCC-3’) 8BF (5’-TGTATGCCAGGACTGCTT-3’), 8BR (5’-GGAGGATTTGTGCA-CAGGACA-3’).

PCR was carried out in a total volume of 15 μL with 50 ng of genomic DNA, 10 nM of each primer, and 9 μl of PCR Master Mix, 2X (Promega, Madison, Wisconsin, U.S.A.). After an initial denaturation step at 95°C for 5 minutes, amplification was performed for 35 cycles (denaturation at 95°C for 1 minute, annealing at 57°C for 1 minute, and extension at 72°C for 2 minutes). PCR products were bidirectionally sequenced using the ABI PRISM BigDye Terminator Cycle Sequencing Ready Reaction Kit and ABI3130 Genetic Analyzer (Applied Biosystems, Foster City, Ca, U.S.A.).

Results

Main results of the coagulation study are summarized in Table 1. Platelets and aPTT were normal. PT was variably prolonged using 6 out of the 7 thromboplastins but was fully corrected by the addition of normal plasma. On the contrary PT was normal using an OX brain thromboplastin. FVII assay was also low with the same six thromboplastins but it was normal with the OX brain preparation. FI, FII, FV, FVII, FIX, FX, F XI, FXII, FXIII were normal in the proposita’s plasma.

<table>
<thead>
<tr>
<th>Test</th>
<th>Proposita</th>
<th>Normal value</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Platelets count</td>
<td>390</td>
<td>150-350 x109/L</td>
<td>Sysmex XT 2000i</td>
</tr>
<tr>
<td>aPTT</td>
<td>26 sec.</td>
<td>26-36 sec.</td>
<td>APTT-SP (Instrumentation Laboratory)</td>
</tr>
<tr>
<td>PT (rabbit brain)</td>
<td>55.8 sec.</td>
<td>14-15 sec.</td>
<td>PT-Fibrinogen HSPlus(Instrument laboratory)</td>
</tr>
<tr>
<td>PT (rabbit brain)</td>
<td>36.9 sec.</td>
<td>12-14 sec.</td>
<td>STA Neoplastin Plus (Stago Laboratory)</td>
</tr>
<tr>
<td>PT (rabbit brain)</td>
<td>83.6 sec.</td>
<td>14-15 sec.</td>
<td>Tromboplastina S (Hemo Medica)</td>
</tr>
<tr>
<td>PT (human placenta)</td>
<td>17.3 sec.</td>
<td>13-15 sec.</td>
<td>Tromborel (Dade-Behring)</td>
</tr>
<tr>
<td>PT (human recombinant)</td>
<td>17.3 sec.</td>
<td>10-12 sec.</td>
<td>Recombiplastin 2G (Instrumentation Laboratory)</td>
</tr>
<tr>
<td>PT (human placenta)</td>
<td>13.1 sec.</td>
<td>10-12 sec.</td>
<td>Innovin Nude (Siemens)</td>
</tr>
<tr>
<td>Ox-brain thromboplastin</td>
<td>40 sec.</td>
<td>38-42 sec.</td>
<td>Thrombostest (Nygaard Laboratories)</td>
</tr>
<tr>
<td>PT × Normal plasma</td>
<td>16 sec.</td>
<td>14-16 sec.</td>
<td>PT-Fibrinogen HSPlus(Instrument laboratory)</td>
</tr>
</tbody>
</table>

**FVII assays:**

| Rabbit brain | 1% | 80-120 | PT-Fibrinogen HSPlus(Instrument laboratory) |
| Rabbit brain | 1% | 80-120 | STA Neoplastin Plus (Stago Laboratory) |
| Rabbit brain | 1% | 80-120 | Tromboplastina S (Hemo Medica) |
| Human placenta | 29% | 80-120 | Tromborel (Dade Behring) |
| OX brain | 96% | 80-120 | Thrombostest (Nygaard Laboratories) |
| Human recombinant | 53% | 80-120 | Recombiplastin 2G (Instrumentation Laboratory) |
| Human recombinant | 39% | 80-120 | Dade Innovin (Siemens) |
| FI, FII, FV, FVIII, FIX, FX, F XI, FXII, FXIII | | 80-120 | |
| Factor VII Antigen | 105% | 80-120 | Asserachrom FVII (Stago Laboratories) |
FII, FV and FX, the other clotting factors that could prolong the test, were normal.

The level of FVII varied between 0.6% to about 96% using the tissue thromboplastins of different origin. It was low with the three rabbit brain thromboplastins (RBT) whereas it was about 40-50% of normal using the human placenta thromboplastin or the two human recombinant thromboplastins and it was 96% of normal using the OX brain preparation. All other clotting factors were within normal limits. Factor VII antigen was 105% of normal.

The daughter had a normal aPTT and a slightly prolonged or borderline PT. FVII level was 37-52% with RBTs and 63-74% of normal with the thromboplastin of human placenta or with the human recombinant ones. Genetic analysis showed that the proposita was homozygote for an Arg304Gln mutation in exon 8 whereas the daughter was heterozygote for the same mutation (Figure 1). The study of the other exons failed to show any other abnormality.

Discussion

The Arg304Gln mutation, together with the Ala294Val represent the most frequent mutations in FVII deficiency [2]. It is interesting to note that, despite the fact that the two mutations are located in the catalytic domain controlled by exons 8, they are different. In fact the Ala294Val mutations is a Type 1 defect (low activity and low antigen) whereas the Arg304Gln (FVII Padua) mutation is a Type 2 defect (low activity, normal antigen) [2,5,6].

Factor VII Padua was first reported in Italy in 1978 on the basis of peculiar clotting tests which showed a discrepancy in the FVII level according to the thromboplastin used in the assay system. Factor VII was low 4-5% of normal using rabbit brain and lung thromboplastin but it was normal when an OX brain thromboplastin was used. Thromboplastins of human origin (placenta) or human recombinant yielded levels of about 30-50% of normal [5,6]. The identity of behavior between human placenta thromboplastin and human recombinant preparation had been established in 1993 [17].

In 1991 it was demonstrated that the defect was due to in Arg304Gln mutation in exon 8 [10,21].

After the first cases, several other patients were described in many parts of the world. A least three main areas of the world show a concentration of the defect, namely the Mediterranean area, the USA (Afro-Americans) and Japan [13,16,18,22].

Since these three areas have nothing in common it is plausible to conclude for a multifounder effect. This indicates that Arg304 is a hot spot for mutations. The description of an Arg304Trp mutation (FVII Nagoya) is in agreement with the hot-spot assumption [23].

The substitution of Arg304 with tryptophan instead of glutamine (FVII Nagoya) does not change the peculiar clotting pattern [23].

Contrary to FVII Padua, FVII Nagoya appears rare, having been described so far only in two patients in Japan [23]. Since the clotting pattern is the same regardless of the mutated glutamine or tryptophan it seems that the Arg304 plays an important role in binding to tissue factor. This is in agreement with the observation that mutation at Pro303Thr is not followed by the appearance of the typical thromboplastin dependent coagulation pattern [24].

These is another area of FVII strictly involved in binding to tissue factor, namely the first EGF domain controlled by exon 4 [25-29].
The exact relation between these two area of FVII involved in binding to tissue factor has not been fully clarified yet [25,26]. It seems that the EGF area (exon 4) mainly binds the protease to its receptor whereas interaction with at least one additional protease domain residue (exon 8) is needed for the full development of the catalytic activity of the bound protease [27].

This apparently is the first homozygote patient with this mutation seen not only in Argentina but also in Latin-America.

A few heterozygotes and a compound heterozygote (combined with the Gly365Cys mutation) have been reported from Brazil, Venezuela and Costa Rica [14,29,30]. These are the only cases listed in FVII mutation Data Bases.

Whether this is due to genetic reasons or to a defective diagnosis remains to be proven. The latter hypothesis seems more likely since this rarity is strange in view of the fact that several cases have been described in European countries which were in the past the source of emigrants to Argentina (mainly Spain, Italy and Israel).

The proposita refers a Spanish background. This is not surprising since occasional patients with a clotting pattern compatible with a FVII Padua diagnosis have been described in Spain [31]. However there is no molecular biology demonstration of the defect. The intense genetic crossing that occurred in the past among the people of the Mediterranean countries and in Argentina herself may fully justify the finding.

There are a few other cases of FVII deficiencies seen in Argentina but they lack molecular biology studies [32-35]. The clotting data available for some of these patients seem to exclude that they were cases of FVII Padua [35]. Due to the paucity or even absence of bleeding symptoms shown by patients with FVII Padua, the administration of aFVII concentrates in the proposita was unjustified. Thrombotic events both arterial and venous have been reported in these patients after replacement therapy [36,37]. Furthermore, when dealing with patients with congenital FVII deficiency, clinical manifestations have always to be kept in mind besides the results of the clotting tests.

A final consideration is indicated. There is today a wide discrepancy between the number of RBD reported in Latin American and a population of about 600 million. Such discrepancy should be eliminated much to the benefit of patients who suffer for a bleeding tendency without knowing the reason for it. Hemophilia A or B and VW Disease patients are probably more easily observed, diagnosed and treated. However even RBD, taken altogether, represent an important segment of the bleeding patients.

We hope this study will stimulate in Argentina and Latin American countries at large, the search for rare bleeding disorders as it has been treated. However even RBD, taken altogether, represent an important segment of the bleeding patients.

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References


